

Antimicrobial Efficacy of Contemporary Obturating Materials used in Primary Teeth- An In-vitro Study

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ABSTRACT

Introduction: Preservation of primary tooth with non-vital pulp has been a major problem for pediatric dentist. In such cases, the preferred treatment modality is pulpectomy. The objective of pulpectomy is complete removal of necrotic and irreversibly infected pulp of an affected tooth but, in spite of chemomechanical preparation and copious irrigation of canal, there are chances of failure of pulp therapy due to the entrapped micro organisms in the canal space owing to the tortuous and complex nature of the root canal system and change in their morphology with root resorption. Thus, for optimal success of endodontic treatment, obturating material with antimicrobial properties is advocated in deciduous teeth.

Aim: The aim of the study was to assess the antimicrobial efficacy of different obturating materials used in pediatric dentistry.

Materials and Methods: In the present study, antimicrobial efficacy of endoflas, Zinc Oxide Eugenol (ZOE), calcium

hydroxide with chlorhexidine, calcium hydroxide with iodoform and distilled water, metapex and saline as control were assessed and compared with each other.

The antimicrobial efficacy of obturating materials was evaluated against *E. faecalis* (ATCC 29212) by agar diffusion method. The statistical analysis was done using SPSS Version 15.0. The Analysis of Variance (ANOVA) and Post-Hoc Tests (Tukey-HSD) were performed to reveal the statistical significance.

Results: Intergroup comparison at 24 hours (Tukey's test) revealed significant difference amongst all the groups. Endoflas had significantly higher zone of inhibition as compared to all the other groups except group treated with ZOE (p-value <0.001).

Conclusion: The antimicrobial activity of obturating materials according to results obtained from the present study can be summarized as follows:

Endoflas > ZOE >Calcium hydroxide + Chlorhexidine > Calcium hydroxide + Iodoform +Distilled water ~ Metapex > Saline.

Keywords: Calcium hydroxide, Chlorhexidine, Endoflas, Obturating materials, Pulpectomy

INTRODUCTION

The primary goal of pulp therapy is to maintain the integrity and health of the teeth and their supporting tissues [1]. The objective of pulpectomy is complete removal of necrotic and irreversibly infected pulp of a tooth affected by caries, traumatic injuries or other causes, so that the tooth remains asymptomatic and functional in the oral cavity till it exfoliates normally [2,3].

Generally, all bacteria that inhabit the oral cavity have the ability to invade the pulp space during and after pulp necrosis in primary teeth. *E.faecalis* being a highly resilient and persistent organism plays an important role in the etiology of recurrent periradicular lesions post endodontic treatment in spite of making a small fraction of the flora in an untreated canal [4]. In spite of chemo mechanical preparation and copious irrigation of canal, there are chances of failure of pulp therapy in primary teeth due to the entrapped micro organisms in the canal space due to the tortuous and complex nature of the root canal system [2,3]. Thus, for optimal success of endodontic treatment, substances with antimicrobial properties are advocated as obturating materials in deciduous teeth. At present, the commonly used materials for obturation of deciduous teeth are Zinc Oxide Eugenol (ZOE), iodoform based pastes, triple antibiotic paste and calcium hydroxide with different vehicles.

ZOE has been traditionally used as an obturating material in the primary dentition [5] and was the first obturating material to be recommended for primary teeth. However, its rate of resorption is slower than that of deciduous teeth [6-8]. Moreover, it can irritate the periapical tissues, can cause necrosis of bone and cementum and may alter the path of eruption of succedaneous tooth [9].

lodoform paste as an obturating material for primary teeth have received attention in the past. Commercially available iodoform based pastes are KRI paste, Maisto paste, Guedes-Pinto paste, Endoflas, Metapex and Vitapex [10,11]. Studies have shown that these iodoform combinations have better resorbability, biocompatibility and antibacterial properties [1,7,12] than ZOE but with a disadvantage that it may produce a yellowish-brown discoloration of tooth crown thereby compromising esthetics [1].

Endoflas is a resorbable paste obtained by mixing a powder containing iodoform (40.6%), zinc oxide (56.5%), calcium hydroxide (1.07%), barium sulphate (1.63%) and liquid consisting of eugenol and paramonochlorphenol.

Calcium hydroxide, which was introduced to dentistry by Hermann in 1930, has been known to promote healing in many clinical situations. It has been used as an obturating material for primary teeth alone and also in association with iodoform [13,14]. Calcium hydroxide can be used with various vehicles such as distilled water, glycerine, chlorhexidine etc.

The present study was undertaken to find out the efficacy of various obturating materials (v.i.z. ZOE, calcium hydroxide with chlorhexidine, endoflas, calcium hydroxide and iodoform in aqueous vehicle, metapex and saline) [Table/Fig-1] used in primary teeth against *E.faecalis*.

MATERIALS AND METHODS

This in-vitro study was carried out in Department of Pedodontics and Preventive Dentistry in collaboration with Department of Pathology and Microbiology, Saraswati Dental College and Hospital, Lucknow, Uttar Pradesh, India, with an overall sample size of 60.

Preparation of broth and bacterial growth: The antimicrobial activity of obturating materials used in primary teeth against E. faecalis (ATCC 29212) was evaluated in this study by agar diffusion method. The standard bacterial strain of *E. faecalis* (ATCC 29212) was obtained from King George Medical University, Lucknow, Uttar Pradesh, India. The purity of test strain was confirmed using the Gram's stain. Only 0.37 grams of Brain Heart Infusion (BHI) broth was added to 10mL of distilled water and mixed by gently shaking the container. This mixture was sterilized by autoclaving for 15 minutes at 121°C and 15lb pressure and then allowed to cool at room temperature. The bacterial strain (E. faecalis ATCC 29212) was inoculated in BHI broth and incubated at 37°C for 24 hours. Following incubation, the cultures were centrifuged at 3000 rpm for 10 minutes. The supernatant liquid was discarded and the precipitate containing microbial cells was separated from the base of test tube. The precipitate containing microbial cells was re-suspended in saline and turbidity of this culture suspension was adjusted until it was equivalent to the no.1 McFarland Standard (Approximately 3 X 10⁸ cells/mL).

S.No	Group	Material	Manufacturer	Composition	Mixing ratio
1.	Group I	Metapex	Meta Biomed Co. Ltd, Korea	lodoform, Calcium hydroxide, Silicone oil	Available in paste form
2.	Group II	Zinc oxide+ Eugenol	Septodont, India Septodont, India	Zinc oxide Eugenol	1:1.5 v/v Liquid powder ratio
3.	Group III	Cavipac + Dentochlor	Pyrex Polykem, India Ammdent, India	Calcium hydroxide Chlorhexidine gluconate (2%)	2gms calcium hydroxide powder per ml of 2% chlorhexidine
4.	Group IV	Endoflas FS	Sanlor lab, Colombia	Powder-Barium sulfate, Zinc oxide, Iodoform, Calcium hydroxide, Liquid-Eugenol, Accelerator-Zinc acetate	Powder and Eugenol was mixed in homogenous thin paste consistency
5.	Group V	Cavipac + lodoform + Distilled water	Pyrex, India Pyrex, India	Calcium hydroxide lodoform Distilled water	Calcium hydroxide: lodoform- 3:1, mixed with Distilled water in paste consistency
6.	Group VI	Normal Saline	Punjab formulations Ltd, Punjab, India	Sodium Chloride- 0.9% w/v (osmolarity-308, Os/L)	

[[]Table/Fig-1]: Obturating materials used in the study.

S No	Group	Description	No. of Samples	Percentage of groupwise distribution of material	
1	I	Metapex	10	16.7	
2	Ш	ZOE	10	16.7	
3	Ш	Ca(OH) ₂ + Chlorhexidine	10	16.7	
4	IV	Endoflas	10	16.7	
5	V	Ca(OH) ₂ + lodoform + Distilled water	10	16.7	
6	VI	Control	10	16.7	
[Table/Fig.2]: Group wise distribution of materials					

Preparation of culture medium: Mueller Hinton agar (15.2gm) was added to 400mL of distilled water and mixed by gently shaking the container. This mixture was sterilized by autoclaving for 15 minutes at 121°C and 15lb pressure. After 15 minutes, the liquid was cooled to room temperature. In a laminar flow chamber, this liquid medium was poured in 20 petri dishes of size 90mm and allowed to set.

Inoculation of bacterial strain on culture media: Each agar plate with 20mL of Mueller Hinton agar was inoculated with 0.1mL microbial suspension using sterile swab. The 3 wells (4mm of depth X 6mm of diameter) were made in each of the agar plates, i.e., total 60 wells were prepared in 20 agar plates to test five different obturating materials and a control.

Placement of obturating materials and incubation: A sterile spatula and glass slab was used for mixing the obturating materials in creamy consistency. Each freshly mixed experimental obturating material was placed in 10 wells of different petri dishes. These agar plates were then incubated at 37°C under anaerobic conditions in an incubator using Microaerophilia (candle jar system). An anaerobic indicator tablet (Anaero indicator tablets) was placed in the jar to monitor oxygen contamination of the environment.

Measuring the size of zone of inhibition: A lack in bacterial colonization was observed for each obturating material. It was indicated by growth inhibitory zones (clearing of agar) around each obturating material. The most uniform diameter segment of the zone of inhibition was determined in millimeters by measuring the shortest distance between the outer margin of the well and initial microbial growth after 24 hours of incubation.

STATISTICAL ANALYSIS

The statistical analysis was done using SPSS Version 15.0. The Analysis of Variance (ANOVA) and Post-Hoc Tests (Tukey-HSD) were performed to know the effect of each variable and to reveal the statistical significance. The values were represented in number(%) and mean±SD.

RESULTS

In the present study, a total of 60 samples were assessed for antimicrobial efficacy against *E. faecalis* [Table/Fig-2]. Antimicrobial efficacy assessment was done at 24 hours time intervals in terms of mean zones of inhibition. At 24 hours, no inhibition was seen in control group. However, in experimental groups mean zone of inhibition was maximum in Group IV (4.53 ± 1.02 mm) followed by Group II (3.80 ± 0.82), Group III (3.10 ± 0.21), Group V (2.10 ± 0.70) and Group I (1.80 ± 0.35) respectively [Table/Fig-3].

Between group comparisons (Tukey HSD test) at 24 hours [Table/ Fig-4], revealed Group IV had significantly higher zone of inhibition as compared to all the other groups except Group II. Group II had significantly higher zone of inhibition as compared to Groups I, V and VI. Group III also had significantly higher mean value as compared to Groups I, V and VI. Group V had significantly higher mean value as compared to Groups I and VI. Group I had significantly higher mean value as compared to Group VI. On the

S No.	Group	Mean	SD	
1	I	1.80	0.35	
2	Ш	3.80	0.82	
3	III	3.10	0.21	
4	IV	4.53	1.02	
5	V	2.10	0.70	
6	VI	0.00	0.00	
F (ANOVA)		66.004		
p-va	lue	<0.001		
[Table/Fig-3]: Intergroup comparison of mean zones of inhibition (in mm) in different				

groups at 24 hours time intervals (ANOVA)

S.No.	Group	Mean Diff.	SE	p-value
1	l vs ll	-2.00	0.28	<0.001
2	l vs III	-1.30	0.28	<0.001
3	l vs IV	-2.73	0.28	<0.001
4	l vs V	-0.30	0.28	0.892
5	l vs VI	1.80	0.28	<0.001
6	ll vs III	0.70	0.28	0.145
7	II vs IV	-0.73	0.28	0.120
8	ll vs V	1.70	0.28	<0.001
9	ll vs VI	3.80	0.28	<0.001
10	III vs IV	-1.43	0.28	<0.001
11	III vs V	1.00	0.28	0.010
12	III vs VI	3.10	0.28	<0.001
13	IV vs V	2.43	0.28	<0.001
14	IV vs VI	4.53	0.28	<0.001
15	V vs VI	2.10	0.28	<0.001

[Table/Fig-4]: Between group comparisons (Tukey HSD test).



basis of above evaluation, the following order of mean zone of inhibition was observed in different groups:

$$\label{eq:constraint} \begin{split} & \mathsf{Endoflas} \geq \mathsf{ZOE} \geq \mathsf{Calcium} \ \mathsf{hydroxide} + \mathsf{Chlorhexidine} > \mathsf{Calcium} \\ & \mathsf{hydroxide} + \mathsf{Iodoform} + \mathsf{Glycerine} \sim \mathsf{Metapex} > \mathsf{Saline}. \end{split}$$

DISCUSSION

The objective of pulp therapy in primary teeth is to preserve the tooth in functional stage until their exfoliation as the teeth act as natural space maintainers, so that the permanent teeth may have adequate space for aligning themselves [15,16]. Also, preservation of primary teeth helps in esthetics, phonetics, mastication and social acceptance. Pulpectomy is indicated when pulp has become irreversibly infected or necrotic due to caries, trauma or other causes [2]. However, tortuous and complex nature of the root canal system and change in their morphology with root resorption makes it difficult to achieve proper cleansing by mechanical instrumentation and irrigation of the canals [2]. For optimal success of pulp therapy, substances with antimicrobial properties are advocated as root filling material in deciduous teeth. Bacterial re-entry and growth of residual micro-organism may be prevented by incorporation of antimicrobial components into obturating materials [17].

In the present study, to assess the antimicrobial activity of endodontic sealers and obturating materials agar diffusion test was employed as it is one of the most commonly used methods [3,13,18]. The agar diffusion method operates on the principle that test material will diffuse from the paper disk or small well into an agar medium containing test organism. It evaluates halos of microbial growth inhibition as reference parameters. This method is advantageous since it directly compares the efficacy of different obturating materials against the micro-organisms thereby determining which material can be used for effectively disinfecting the pulp canal space [19]. However, the results can be influenced by the degree of diffusion of material across the medium [19].

Success rate of root canal treatment can be lowered significantly if there is presence of *E.faecalis* in pulp canal system at the time of obturation [11]. Most importantly, *E.faecalis* efficiently and rapidly colonizes dentinal tubules compared to other species and therefore is particularly difficult to eradicate [20]. It represents a standard against which the antimicrobial action of a medicament should be tested [20,21].

The present study was undertaken to assess the antimicrobial activity of different obturating materials used for primary teeth. For this purpose, five combinations of obturating materials and saline as control were analysed against *E.faecalis* by agar diffusion method. The data was collected and statistically analysed for intergroup and between group comparisons (Tukey HSD test) changes at 24 hours time intervals.

In the present study, endoflas, calcium hydroxide with chlorhexidine, fresh mix of calcium hydroxide with iodoform using distilled water as a vehicle were tested as there is very sparse literature and studies present regarding them, as an obturating material in primary teeth, whereas ZOE and metapex are regularly used in dental practice for obturation of primary teeth.

On intergroup comparison, the results revealed that all the materials tested possessed antimicrobial activity as demonstrated by the formation of growth inhibition zones. Endoflas showed maximum mean zone of inhibition against *E.faecalis* followed by ZOE and combinations of calcium hydroxide (calcium hydroxide + chlorhexidine, calcium hydroxide + iodoform + distilled water, metapex) [Table/Fig-5,6].

Eugenol based root canal obturating material (Endoflas and ZOE) showed highest antimicrobial activity as compared to the non-eugenol containing in the present study. The antimicrobial effect of eugenol based obturating material was mainly attributed to the action of eugenol. Eugenol acts on micro-organism by causing protein denaturation rendering it non-functional [22-24]. The results of the various studies performed by Gomes et al., Markowitz et al., and Saggar et al., also confirmed that eugenol containing obturating materials were more superior in inhibiting the microorganisms [22,24,25].

Endoflas FS showed marginally better, antimicrobial activity than ZOE obturating material probably due to incorporation of known bactericidal agents such as iodoform [22]. Iodoform acts by the liberation of iodine. It is believed that iodine, which is an oxidizing agent, can irreversibly oxidize and thus, inactivate essential metabolic compounds like proteins, nucleotides and fatty acid resulting in cell death, but the exact mode of action is not fully known [25]. Similar results have been reported by Gopikrishna et al., and Kayaoglu et al., [23,26].

In the present study, it was also observed that various calcium hydroxide combinations showed inhibitory action against *E. faecalis*. Calcium hydroxide and iodoform when mixed with aqueous vehicle performed better than metapex that contain silicone oil as a vehicle. It has been asserted that all biological actions of calcium hydroxide are due to the ionic dissociation in Ca²⁺ and OH⁻ ions. The vehicle in calcium hydroxide formulations plays an important role in the overall process because it determines the velocity of ionic dissociation. Calcium hydroxide, when mixed with aqueous vehicle, allows better ionic dissociation and hence, diffusion and greater zone of inhibition in the agar than when mixed with oily vehicle [26].

Among the different preparations of calcium hydroxide, calcium hydroxide with chlorhexidine showed the best result followed by freshly prepared mix of calcium hydroxide and iodoform while metapex showed minimum inhibitory action. The possible reason could be that, chlorhexidine gluconate is a broad spectrum antimicrobial agent and is effective against bacteria and fungi [27-31]. Thus, antimicrobial activity of calcium hydroxide increases when mixed with chlorhexidine [16]. Pabla et al., in their study reported lesser antimicrobial activity of metapex against aerobic and anaerobic bacteria in comparison to ZOE, KRI paste and MAISTO paste [16]. However, Garcia-Godoy and Nurko studied the effectiveness of metapex in the root canal treatment of primary teeth and reported treatment to be deemed successful as clinically the tooth was painless, with no pathological mobility, healthy gingiva and no fistulation [1,13].

It was observed from the present study that eugenol based root canal obturating material particularly endoflas showed the highest antimicrobial activity against *E.faecalis* at 24 hours time intervals.

The rationale for performing this invitro study is to offer information to clinicians about antimicrobial efficacy of different root canal obturating materials used in primary tooth against E.faecalis.

LIMITATION

Limitation of this study could be that the antimicrobial efficacy of obturating materials has been evaluated in in-vitro conditions which may be modified in a clinical setup due to presence of dentin and serum. Hence, further in-vivo studies with larger sample size are needed to evaluate the antimicrobial efficacy of root canal obturating materials, in clinical settings.

CONCLUSION

Within the limitations of the study, all materials produced zones of microbial growth inhibition against the tested microorganism E. faecalis. On intergroup comparison of mean zones of inhibition in different groups at different time intervals, endoflas showed maximum mean zone of inhibition followed by ZOE, various combinations of calcium hydroxide and metapex showed least antimicrobial activity. The mean zones of inhibition can be summarised as follows:

Endoflas > ZOE > Calcium hydroxide + Chlorhexidine > Calcium hydroxide +lodoform +Distilled water ~ Metapex > Saline.

REFERENCES

- [1] Garcia-Godoy F. Evaluation of an iodoform paste in root canal therapy for infected primary teeth. ASDC J Dent Child. 1987;54(1):30-34.
- [2] American Academy of Pediatric Dentistry. Guideline on pulp therapy for primary and young permanent teeth. Pediatr Dent. 2009;31(6):179-86.
- Bodrumlu E, Semiz M, Antibacterial activity of a new endodontic sealer against [3] Entercoccus faecalis. J Can Dent Assoc. 2006;72:637.
- [4] Rifkin A. The root canal treatment of abscessed primary teeth: A three to four year follow-up. J Dent Child. 1982;49:428-31.
- [5] Am Dent Assoc. 1930;17:1150–53.
- Tchaou WS, Turng BF, Minah GE, Coll JA. Inhibition of pure cultures of oral [6] bacteria by root canal filling materials. Pediatr Dent. 1996;18(7):444-49.
- [7] Mortazavi M, Mesbahi M. Comparison of zinc oxide and eugenol, and vitapex for root canal treatment of necrotic primary teeth. Int J Paediatr Dent. 2004;14(6):417-24.

- [8] Zalp NO, Aroglu IS, Sonmez H. Evaluation of various root canal filling materials in primary molar pulpectomies: an in vivo study. Am J Dent. 2005;18(6):347-50.
- [9] Grossman L.I. Microbiology. Chapter 2 In: Grossman's Endodontics Practice. 12th ed. Chandra B.S., Krishna V.G, editor. Lippincott William & Wilkins;2010.p.43-52.
- [10] Chawla H, Setia S, Gupta N, Gauba K, Goyal A. Evaluation of a mixture of zinc oxide, calcium hydroxide, and sodium fluoride as a new root canal filling material for primary teeth. J Indian Soc Pedod Preven Dent. 2008;26(2):53-58.
- [11] Kriplani R, Thosar N, Baliga MS, Kulkarni P, Shah N, Yeluri R. Comparative evaluation of antimicrobial efficacy of various root canal filling materials along with aloevera used in primary teeth: A microbiological study. J Clin Pediatr Dent. 2013;37(3):257-62.
- [12] Primosch RE. Primary tooth pulp therapy as taught in predoctoral pediatric dental programs in the United States. Pediatr Dent. 1997;19(2):118-22.
- [13] Nurko C, Ranly DM, Garcia Godoy. Resorption of a calcium hydroxide/ iodoform paste (vitapex) in root canal therapy for primary teeth: a case report. Pediatr Dent. 2000;22:517-20.
- [14] Chawla HS, Mani SA, Tewari A. Calcium hydroxide as a root canal filling material in primary teeth-a pilot study. J Indian Soc Pedod Preven Dent. 1998;16(3):90-92.
- [15] Weiss El, Shalhav M, Fuss Z. Assessment of antibacterial activity of endodontic sealers by a direct contact test. Dent Traumatol. 1996;12(4):179-84.
- [16] Pabla T, Gulati MS, Mohan U. Evaluation of antimicrobial efficacy of various root canal filling materials for primary teeth. J Indian Soc Pedod Prev Dent. 1997;15:134-140.
- [17] Smadi L, Mahafzah A, Khraisat A. An in vitro evaluation of antimicrobial activity of nine root canal. J Contemp Dent Pract, 2008; 5:1-8.
- [18] Estrela C, Bammann LL, Pimenta FC, Pécora JD. Control of microorganisms in vitro by calcium hydroxide pastes. Int Endod J. 2000;34:416-18.
- Estrela C, Estrela CRA, Bammann LL, Pécora JD. Two methods to evaluate the [19] antimicrobial action of calcium hydroxide paste. J Endod. 2001;27:5-7.
- [20] Marsh SJ, Largent MD. A bacteriologic study of the pulp canals of infected primary molars. ASDC J Dent Child. 1967;34:460-70.
- [21] Cohen MM, Joress SM, Calisti LP. Bacteriologic study of infected deciduous molars. Oral Surg Oral Med Oral Pathol. 1960;13:1382-86.
- [22] Gomes BPFA, Pedroso JA, Jacinto RC, Vianna ME, Ferraz CC, Zaia AA, et al. In vitro evaluation of the antimicrobial activity of five root canal sealers. Braz Dent J. 2004; 15:30-35.
- [23] Gopikrishna AV, Kandaswamy D, Jeyaval Rajan K. Comparative evaluation of the antimicrobial efficacy of five endodontic root canal sealers against enterococcus faecalis and candida albicans. J Cons Dent. 2006;9(1): 2-11.
- [24] Markowitz K, Moynihan M, Liu M, Kim S. Biologic properties of eugenol and zinc oxide eugenol and zinc oxide eugenol. A clinically oriented review. Oral Surg Oral Med Oral Pathol. 1992;73:729-37.
- Saggar V, Chandra S, Jaiswal JN, Singh M. Antimicrobial efficacy of iodoformized [25] zinc oxide eugenol sealer on microorganisms of root canal. J Ind Soc Pedod Preven Dent.1996;14(1):1-3.
- [26] Kayaoglu G, Orstavik D. Virulence factors of Enterococcus faecalis: relationship to endodontic disease. Crit Rev Oral Bio Med. 2004;15(5):308-20.
- [27] Neelakantan P, Sanjeev K, Subbarao CV. Duration- dependent susceptibility of endodontic pathogens to calcium hydroxide and chlorhexidine gel used as intracanal medicament: an invitro evaluation. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2007;104:138-41.
- [28] Pabla T, Gulati MS, Mohan U. Evaluation of antimicrobial efficacy of various root canal filling materials for primary teeth. J Indian Soc Pedod Prev Dent. 1997:15:134-140.
- Estrela C, Pécora JD, Souza-Neto M.D, Estrela CRA, Bammann LL. Effect of [29] vehicle on antimicrobial properties of calcium hydroxide pastes. Braz Dent J. 1999;10:63-72
- [30] Basrani B, Tjäderhane L, Santos JM, Pascon E, Grad H, Lawrence HP, et al. Efficacy of chlorhexidine and calcium hydroxide containing medicaments against Enterococcus faecalis in vitro study. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2003;96:618-24.
- Rewal N, Thakur AS, Sachdev V, Mahajan N. Comparison of endoflas and zinc [31] oxide eugenol as root canal filling materials in primary dentition. J Ind Soc Pedod Preven Dent. 2014;32(4):318-21.

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