

Surgical Site Infection Caused by *Aeromonas hydrophila* in a Patient with Underlying Malignancy

FRINCY KHANDELWAL BARUAH¹, NISHAT HUSSAIN AHMED², RAJESH KUMAR GROVER³

ABSTRACT

Aeromonas skin and soft tissue infections in cancer patients can lead to serious life threatening conditions such as cellulitis, necrotizing fasciitis and myonecrosis. We report here a case of surgical site infection, post radical mastectomy, in a 58-year-old female with carcinoma breast. Cultures of exudates from the wound grew *Aeromonas hydrophila* on repeated occasions. Recovery was uneventful following targeted antimicrobial therapy and regular dressing of the wound. Early suspicion, diagnosis, and treatment with potent antibiotics are needed to prevent any further complications resulting from infection by this emerging pathogen.

Keywords: Carcinoma breast, Immunocompromised, Necrotising fasciitis, Radical mastectomy, Surgical site infection

CASE REPORT

A 58-year-old female, admitted to Delhi State Cancer Institute, a tertiary care centre in Delhi on 23rd Dec'2013 with carcinoma breast, underwent modified radical mastectomy. Postoperative stay was uneventful and the patient was discharged from the hospital in satisfactory condition. However, after two weeks, she revisited with complaints of pain, redness and purulent discharge from the site of surgery. Gram stain of the purulent material revealed plenty of polymorphonuclear cells, and no microorganisms. Later, on culture, pure growth of non-lactose fermenting colonies was obtained on MacConkey agar while on blood agar, the colonies were small, round, opaque and beta hemolytic [Table/Fig-1a&b]. The organism was found to be a Gram negative bacillus and motile. The colonies were catalase test and oxidase test positive. The Gram negative rods were identified as *Aeromonas hydrophila* by the Vitek-2 compact system (BioMe'rieux, North Carolina/USA) with Gram negative GN REF21341 identification (GNID) card. Antimicrobial sensitivity was performed using AST-N281 card which showed the organism to be sensitive to piperacillin/tazobactam (MIC \leq 4 μ g/ml), ceftazidime (MIC \leq 1 μ g/ml), cefepime (MIC \leq 1 μ g/ml), cefoperazone/sulbactam (MIC \leq 8 μ g/ml), ciprofloxacin (MIC \leq 0.25 μ g/ml), levofloxacin (MIC \leq 0.12 μ g/ml), trimethoprim/sulfamethoxazole (MIC \leq 20 μ g/ml), imipenem (MIC \leq 0.25 μ g/ml) and meropenem (MIC \leq 0.25 μ g/ml). A second pus sample from the same site also yielded *Aeromonas hydrophila* with the same antibiogram which confirmed the pathogenicity of this rare organism. Blood culture obtained on this occasion was found to be sterile. Daily dressing of the wound and administration of levofloxacin, 500 mg for five days helped in proper healing of the wound. Follow up after two weeks was satisfactory and the wound became clean with healthy granulation tissue.

DISCUSSION

Aeromonads are Gram-negative rods, oxidase-positive, facultatively anaerobic, widely distributed in aquatic environment [1]. Typically, patients acquire *Aeromonas* species infection by oral consumption or direct contact with contaminated water or sea food [2].

Aeromonas infection is often polymicrobial and fatality rates range from 28% to 46% in cases of bacteremia mostly caused by *Aeromonas hydrophila* and *Aeromonas veronii* biovar *sobria* [2,3].

In immunocompromised patients, serious fatal infections such as hepatobiliary infection, invasive skin and soft tissue infections, primary bacteremia, burn infections, pleuropulmonary infection, meningitis and endocarditis may also occur [2,4].

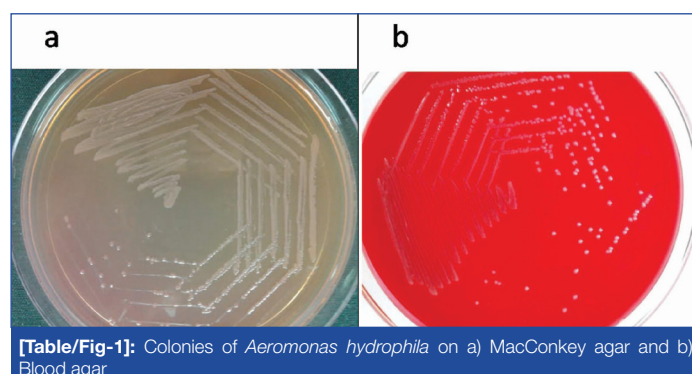
Skin and soft tissue infection by *Aeromonas hydrophila* has been associated with a number of complications such as abscesses, fatal myofascial necrosis, septic arthritis, septic shock, gas gangrene and extremity amputation [5,6].

Necrotizing fasciitis or myonecrosis is often seen in patients with underlying malignancy and can be associated with high mortality rates approaching 60% to 75% [4,7].

In the present case, the patient presented with a non-healing ulcer at surgical site post radical mastectomy which was infected with *A. hydrophila*. Infection in this case may have been acquired from exposure of wound to contaminated water as this organism has been reported to be present in aqueous environments [1].

An earlier study reported *A. hydrophila* to be the most common species of *Aeromonas* causing skin and soft tissue infection among cancer patients [3]. Papadakis et al., reported a patient with fulminant soft tissue infection and septicemia caused by *A. hydrophila* during acute lymphoblastic leukemia treatment who recovered well with antibiotic administration and surgical management [8]. Kuo chin et al., also reported a case of serious necrotizing fasciitis due to *A. hydrophila* in an immunocompromised host which resulted in death of the patient [5].

In India, Mukhopadhyay et al., reported eight cases of *Aeromonas* infection; out of which, there was one case of skin and soft tissue infection due to *A. hydrophila* in a 55-year-old male with underlying soft tissue sarcoma. The patient had recovered well with antibiotics and surgical excision of the intramuscular mass [9]. Sood et al., reported isolation of *Aeromonas hydrophila* from necrotic tissue obtained during fasciotomy in an immunocompetent patient which was resistant to carbapenems and all second and third generation cephalosporins. The patient died within two days of hospitalization due to sepsis and multi organ failure [10].



[Table/Fig-1]: Colonies of *Aeromonas hydrophila* on a) MacConkey agar and b) Blood agar

Though *Aeromonas* isolates are susceptible to a broad range of antibiotics, the members of this genus may also produce beta lactamase which makes them resistant to ampicillin and first-generation cephalosporins.

Timely diagnosis and prompt antimicrobial administration helped in complete recovery of our patient.

CONCLUSION

Thus, in light of the complications associated with *Aeromonas* skin and soft tissue infection, utmost microbiological vigilance is required to promptly identify this rare organism, especially in immunocompromised patients such as those with underlying malignancy; so that timely management of the condition is possible with appropriate antibiotics and surgical debridement, irrigation etc, if required.

REFERENCES

- [1] Igbinosa IH, Igumbor EU, Aghdasi F, Tom M, and Okoh AI. Emerging *Aeromonas* species infections and their significance in public health. *Scientific World Journal*. 2012;2012:625023.
- [2] Okumura K, Shoji F, Yoshida M, Mizuta A, Makino I, Higashi H. Severe sepsis caused by *Aeromonas hydrophila* in a patient using tocilizumab: a case report. *J Med Case Reports*. 2011;5:499.
- [3] Chao CM, Lai CC, Gau SJ, Hsueh PR. Skin and soft-tissue infection caused by *Aeromonas* species in cancer patients. *Journal of Microbiology, Immunology and Infection*. 2013;46 (2):144-46.
- [4] Janda JM, Abbott SL. The genus *Aeromonas*: taxonomy, pathogenicity, and infection. *Clin Microbiol Rev*. 2010;23:35-73.
- [5] Kuo-Chun L, Po-Tsung Y, Cheng L. Necrotizing fasciitis caused by inconspicuous infection of *Aeromonas hydrophila* in an immunocompromised host. *J Surg Case Rep*. 2010;7:2.
- [6] Behera B, Bhorawal S, Mathur P, Sagar S, Singhal M, and Misra MC. Post-traumatic skin and soft tissue infection due to *Aeromonas hydrophila*. *Indian J Crit Care Med*. 2011;15(1):49-51.
- [7] Fatima A, Afridi FI, Qureshi A, Farooqi BJ, Hussain A. Bacteremia due to *Aeromonas hydrophila* in Acute Lymphoblastic Leukemia. *J Coll Physicians Surg Pak*. 2013;23(12):893-95.
- [8] Papadakis V, Poniros N, Katsibardi K, Charissiadou AE, Anastosopoulos J, Polychronopoulou S. Fulminant *Aeromonas hydrophila* infection during acute lymphoblastic leukemia treatment. *J Microbiol Immunol Infect*. 2012;45:154-57.
- [9] Mukhopadhyay C, Chawla K, Sharma Y, Bairy I. Emerging extra-intestinal infections with *Aeromonas hydrophila* in Coastal region of Southern Karnataka. *J Postgrad Med*. 2008;54:199-202.
- [10] Sood S, Nerurkar V. Fatal Necrotizing Soft Tissue Infection by *Aeromonas hydrophila*. *Journal of Clinical and Diagnostic Research*. 2014;8(4):6-7.

PARTICULARS OF CONTRIBUTORS:

1. Senior Resident, Department of Laboratory Medicine, Delhi State Cancer Institute, Delhi, India.
2. Assistant Professor, Department of Laboratory Medicine, Delhi State Cancer Institute, Delhi, India.
3. Director and Chief Executive Officer, Department of Clinical Oncology, Delhi State Cancer Institute, Delhi, India.

NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Dr. Nishat Hussain Ahmed,
Assistant Professor, Department of Laboratory Medicine, Delhi State Cancer Institute, Delhi- 110095, India.
E-mail : drnishathussain@rediffmail.com

FINANCIAL OR OTHER COMPETING INTERESTS: None.

Date of Submission: **Sep 02, 2014**

Date of Peer Review: **Sep 12, 2014**

Date of Acceptance: **Oct 31, 2014**

Date of Publishing: **Jan 01, 2015**