Role of Mycobacterial Culture and Drug Sensitivity Testing Laboratory under National Tuberculosis Elimination Program for the Abolition of Tuberculosis in India by 2025

NANDINI SINGH¹, AMRESH KUMAR SINGH², SUSHIL KUMAR³, NARENDRA PRATAP SINGH⁴, VIVEK GAUR⁵

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ABSTRACT

Microbiology Section

India has made a bold promise to eradicate Tuberculosis (TB) by 2025 five years ahead of the global target. Although, onefourth of the global burden with highest new cases of TB is shown by the country. So yet, no comprehensive analysis has been published on India's National Tuberculosis Elimination Program (NTEP) (2017-2025). The current review details the advanced diagnostic methods like Fluorescence Microscopy (FM), culture, nucleic acid amplification test (Cartridge Based Nucleic Acid Amplification Test (CBNAAT) and True Nucleic Acid amplification Test (TrueNAT)) and Line Probe Assay (LPA) as well as the role and network of mycobacterial Culture and Drug Sensitivity Testing (CDST) laboratories in national scaling-up of evidence-based policies and facilities, which is a critical component in India's fight against TB. The material of this study was mostly obtained from policy and program making documents of World Health Organisation (WHO) and annual TB reports of India. India's TB annual report 2021 says that only half of the patients were successfully treated in the period of conventional longer care regimens. The interventions to achieve the factors related patient's care have been implemented through universal drug sensitivity testing through CDST laboratories, which has driven therapy with a shorter regimen, newer medications, and social protection. In one hand, the comprehensive monitoring scheme through CDST laboratories for TB including all possible drug-resistance cases and other hand, patient's systemic treatment through shorter, more reliable and safer first- or second-line drug regimens are all necessary milestones to achieve the goal of our government for abolition of TB in India by 2025.

Keywords: Cartridge based nucleic acid amplification test, Culture and drug sensitivity testing, Drug resistant tuberculosis, Line probe assay, True nucleic acid amplification test

INTRODUCTION

Tuberculosis is an infectious disease mainly caused by bacteria known as Mycobacterium tuberculosis Complex (MTBC) [1]. Inspite of hard efforts taken to control TB by government and social platforms, this disease continues to be one of the major public health problems worldwide, particularly in developing countries [2]. According to WHO, an estimated 10 million people fell ill with TB altogether over the world which incorporates 5.6 million men, 3.2 million women and 1.2 million children among these India shares up to 25% of total cases [3]. TB tends to spread rapidly due to their asymptomatic existence including lack of early and reliable diagnosis become responsible for higher rate of morbidity and mortality. National services may do a better job of integrating existing diagnostic tools, but new and better tools are needed to allow low-cost, rapid and reliable TB screening closer to the point of treatment, as well as to ensure that all people at risk of TB receive the care they need [4].

The MTBC might be resistance against first line antitubercular drugs either isoniazid and rifampicin or both recognised as Multidrug Resistance (MDR-TB), however; resistance with not only by rifampicin and isoniazid but also with any fluoroquinoline (FQ) including any second line injectable drugs are counted as Extensively Drugresistance (XDR-TB) [5,6]. Drug-Resistant Tuberculosis (DR-TB) has posed a relentless threat to successful TB control. Its existence has been recognised since the first anti-TB medicines were developed for the treatment of TB [7]. The introduction of MDR-TB and more recently XDR-TB has highlighted the need of possible required advanced DST and new medicines or their alternatives for their elimination [8]. To prevent the spreading of DR-TB, early and appropriate diagnosis and complete treatment with less Turnaround Time (TAT) is required [9]. Early detection has increased the mapping of high-risk populations and carefully designed systemic surveillance for active disease among them, which can be helpful to minimise the MTBC infection [10].

India has already taken many crucial measures with impressive and visionary policies in recent years to place itself as a pioneer for a TB-free nation [11-12]. Firstly, Government of India launched National Tuberculosis Programme (1962) followed by pilot programme Revised National TB Control Programme (RNTCP) in 1993 and fully launched 1997, now known as NTEP in 2020 with the aim of making India a TB-free country up to 2025 [13-15]. It is a remarkable and optimistic goal; nevertheless, achieving this status would require implementation of massive and large-scale diagnostic and treatment policies. It functions as a flagship component of the National Health Mission (NHM) and provides technical as well as managerial leadership to anti-TB activities within the country [16]. However, to tackle the large or undiagnosed issues, especially MDR and XDR-TB cases, the country requires high quality medical laboratories that can facilitate not only the diagnosis but also the drug sensitivity testing by covering the maximum cases throughout the endemic area of TB. Therefore, frequent diagnosis and proper treatment of DR-TB remains a major priority of our public health programme. The aim of this review was to summarise the NTEP endorsed different diagnostic methods and their role and laboratory networks towards the abolition of TB from India.

SELECTION OF REVIEWS

This review was based on information published on WHO annual TB reports as well as policy and programme documents of Central TB Division, Ministry of Health and Family Welfare, India [3,17].

For evaluating the efficiency and sensitivity of CDST laboratories for evaluation of *Mycobacterium* either it is genotypic or phenotypic, authors found out 35 article from PubMed and Google scholar as searched term "mycobacterial culture and drug sensitivity testing laboratory AND India". However, only 15 articles were present in internet and found suitable for CDST laboratories evaluation after thorough review. Authors have evaluated further 12 articles from google scholar for information about composition of media for solid and liquid culture and also adopted from guidelines given by WHO [18,19]. Definitions of different diagnostic methods were taken from textbooks of microbiology and information about genotypic technology TrueNAT was taken from Indian Council of Medical Research (ICMR) guidelines [20,21].

MAJOR DIAGNOSTIC METHODS UNDER NTEP

Molecular assays based on nucleic acid amplification techniques have been developed for the fast, sensitive, and reliable diagnosis of TB with the potential to determine their drug susceptibility status simultaneously [22]. Although, NHM provide the diagnostic services through a network of various types of laboratories in three tier fashion under the umbrella of NTEP [23]. In which, they constituted by facilities of microscopy, CDST like solid Lowenstein-Jensen (LJ media) and liquid culture (MGIT960), CBNAAT and TrueNAT and rapid molecular tests like LPA [10]. Their sensitivity, specificity, advantage and disadvantages and descriptions of these diagnostic methods are briefly discussed in [Table/Fig-1] [15,24].



[Table/Fig-1]: Mycobacterial culture and drug sensitivity testing (C&DST) laboratories based advanced techniques for detection of *M. tuberculosis* under National Tuberculosis Elimination Program (NTEP) [15,24].

(A) Fluorescence Microscopy (FM)

Microscopic analysis of clinical sputum specimens has been the major part of TB diagnosis over a century [25]. FM of sputum smear has been used to improve the sensitivity as compared to traditional Zeihl Neelsen (ZN) microscopy [26]. Direct microscopic analysis is frequently used method of TB diagnosis in low income countries like India. Sputum microscopy especially FM is not only affordable for diagnosis but also the determination of reaction to treatment of TB [27]. It retains the primary stain even after decolourisation as well as counter stain to highlight the MTBC for easier recognition [28]. In this technique, the use of Light Emitting Diode (LED)-FM is very helpful for the identification of smear-positive cases of MTBC among the heavy loaded Direct Microscopic Centres (DMCs) and medical colleges under the CDST laboratories [29]. The WHO reviewed the evidence for LED microscopy's effectiveness in 2009, using criteria suitable for assessing both the efficacy and the impact of new TB diagnostics on patients and public health [30]. LED-FM microscope is cost-effective, uses less energy and can be powered by batteries; additionally, the bulbs have a longer life span without harmful compounds if destroyed.

(B) Solid Culture Lowenstein-Jensen (LJ) Media

Solid culture media like LJ medium is a conventional method for CDST; it is less expensive and more readily available than other techniques. It is the most often used medium for culturing the MTBC recommended by the International Union against Tuberculosis [31]. It shows improved sensitivity over the smear with detection limit 100 bacilli/mL [32]. It is mainly composed of malachite green, glycerol and coagulated egg, in which, potassium dihydrogen phosphate anhydrous (KH_2PO_4), magnesium sulphate (MgSO_4.7H₂O) and magnesium citrate are also found in LJ medium to prevent gram positive and gram negative bacteria from growing and limiting growth to only *Mycobacterium* [18]. However, other bacteria are inhibited by the presence of malachite green in the medium. LJ culture showed weak growth rate of MTBC, at least 6-10 weeks for incubation including taken too much time for the result in comparison to liquid culture, is the limitation of this technique [20].

(C) Liquid Culture Media (BACTEC 960)

The BACTEC Mycobacterial Growth Indicator Tube (MGIT) 960 is a gold standard liquid culture method which is used as an invitro diagnostic instrument and resource-constrained environment as recommended by WHO [33-35]. It has been also approved for *M. tuberculosis* diagnosis in DST under the NTEP [24]. The MGIT 960 culture tubes contain 7 mL of Middlebrook 7H9 broth base, to which an enrichment supplement was added according to the instructions of the manufacturer, as well as mixture of antibiotics (MGIT PANTA) consisting of polymyxin B, amphotericin B, nalidixic acid, trimethoprim, and azlocillin [36]. It is used especially for phenotypic diagnosis and drug susceptibility testing of TB, not only just for first-line drugs but also the second-line drugs among many laboratories of DST [19].

(D) Nucleic Acid Amplication Test (CBNAAT and TrueNat)

Nucleic Acid Amplication Test (NAAT) is offered the diagnosis of TB among children, high-risk population living with Human Immunodeficiency Virus (HIV) and extrapulmonary TB cases and also very useful among patients with TB who showed smear-negative through X-ray and preferable cases referred from private sector for early detection and appropriate treatment [37]. CBNAAT is an automatic cartridge-based molecular technique that detects MTBC as well as rifampicin resistance within two hours. It has been endorsed by WHO as an initial diagnostic test among patients suspected with MDR-TB in both pulmonary and extrapulmonary cases [38]. Unlike traditional NAATs techniques, CBNAAT or Xpert MTB/RIF cartridge show sample processing combines with Polymerase Chain Reaction (PCR) amplification and identification through single self-contained research device [38]. This assay covers the significant step forward platform and versatile tool for early diagnosis among all type of DR-TB cases in the fight against TB [39]. In other hand, chip based advanced technique like TrueNAT was developed by joint venture of Bigtec Laboratories and research and development subsidiary section of Molbio Diagnostics [40]. TrueNAT is cost effective when deployed at Point Of Care (POC) and also it is replacing smear microscopy as it can detect the cases more correctly [41]. This technique uses the real-time micro PCR technology and functional among wide range of environmental conditions with minimal user input in primary healthcare settings [21].

(E) Line Probe Assay (LPA)

The LPA is a reverse hybridisation procedure in which the patient's sample is hybridised with membrane strips coated with complementary markers of individual genes [42]. It is a group of Deoxyribonucleic Acid (DNA) strip-based tests that evaluate the MTBC strain's drug resistance profile. It work by examining the amplicons bind to wild-type DNA series probes that target the most common resistance related mutations to first- and second-line agents [31,43]. It also generates results very fast within 24-48 hours [44]. They can detect anti-tubucular drugs resistance status of both isoniazid and rifampicin through the identification of mutation in the

rpoB, katG, and *inhA* genes [45]. In 2008, WHO approved the use of commercial LPAs for detecting MTBC in sputum smear positive specimens (direct testing) and cultured isolates of MTBC with drug resistance specimens (indirect testing) [44]. Using LPAs in countries with a high MDR/XDR strain allows for adequate, prompt care, lowering delivery speeds, morbidity and improving patient outcomes [46]. In recent advances among drug susceptibility testing, WHO also recommended this technique as initial test for second line-LPA for fluoroquinolones and injectable drugs resistance detection, instead of phenotypic culture [44]. Now, many NTEP laboratories use this technique for accurate and fast molecular DST assay for MDR and XDR-TB.

Role of Mycobacterial Culture and Drug Sensitivity Test (CDST) Laboratories

Laboratories are essential for monitoring the diagnosis and treatment of TB. Many laboratory techniques are used in detection especially microbial agent separation, causative bacteria, and drug susceptibility testing of isolates [47]. It becomes increasingly complex with the expansion of quality assured smear microscopy and novel CDST laboratory tools e.g., LPA and CBNAAT [43]. Since, multiple methods are needed to recover, classify and assess drug resistance for MTBC for the confirmation of any single case of TB [25]. Especially, treatment of MDR-TB is a difficult task that should be performed by qualified physicians in centres with reliable mycobacterial culture and in-vitro sensitivity testing services [48].

As per WHO hierarchical management system, NTEP play a significant role of Mycobacterium CDST laboratories as well as quality assurance organisation from the highest level of National Reference Laboratories (NRL) followed to State Intermediate Reference Laboratories, district/ subdistrict level, and finally up to peripheral level of microscopy centres launched by Government of India [10]. Building on this vast laboratory network, Universal Drug Susceptibility Testing (UDST) was introduced freely or less expensive to patients for all type of drug resistance testing throughout the country. However, 100% CDST based quality assured laboratories with efficient capacity and timely identification of patients is the need of hour [49]. National Strategic Plan (NSP) (2017-2025) advocates the early identification of presumptive patients at the first POC among private or public sectors and highly sensitive diagnosis to provide the universal access of TB including DR-TB throughout the country [12,50]. If we see the India TB report 2021, conventional drug susceptibility testing of MTBC with liquid medium is well established and offers time saving and reliable results against a variety of first line and second line antituberculosis drugs [35]. Patients with high-risk of MDR-TB are diagnosed using WHO endorsed rapid diagnostics like CBNAAT/LPA/TrueNAT. However, response to treatment for MDR is always monitored by follow-up on liquid culture (MGIT960) system. Mostly laboratories including our institute Baba Raghav Das Medical College, Gorakhpur, India, also performed commercial Immunochromatic Test (ICT) for identification of Mycobacterium species in all detected cases [10].

All the patients on drug regimen also require TB culture because it helps to check whether the patient is taking his medicine in continuation or not. Since, no laboratory diagnostic is 100% full proof and also molecular detection depends on the presence of resistance conferring mutation. So, development of any new mutation cannot be detected by genotypic methods only and therefore, there is also need of phenotypic tests to identify drug resistance [23]. Modern techniques in laboratories for CDST like liquid culture or LPA etc can make our pathway easier towards the elimination of TB. It is a cost-effective and time-saving means of detecting MDR-TB, as well as a life-saving technique for early identification and treatment [6]. However, LPAs may not reduce the need for traditional CDST capabilities, as culture is still needed for conclusive TB diagnosis in smear negative patients, and DST is required for validation, if MDR/XDR-TB is not diagnosed [18]. Historically, MTBC was identified by phenotypic methods, such as morphological characteristics, growth rates, preferred growth temperature, pigmentation and series of biochemical tests. New phenotypic and genotypic susceptibility testing approaches includes the appealing of both first and second line drugs. Total 28,58,713 tests were performed by the CBNAAT and 1,25,923 tests by TrueNAT in which 53826 (7%) and 340 (3.1%) cases of DR-TB respectively among total confirm TB cases tested [Table/Fig-2]. The first line LPA detected 7.5% cases of MDR-TB and second line LPA detected 5.80% XDR-TB among total confirm TB cases tested. Total cases conducted by liquid culture are 2,85,775 and this second line DST detected approximately 5.2% cases of XDR-TB among 10184 cases [17]. The TAT has been further reduced by molecular detection of drug resistance and appears lower cost of testing followed to become the future of TB diagnosis in all the settings.

Genotypic diagnostic techniques	CBNAAT	Second line liquid culture	First line LPA	Second line LPA	TrueNAT
Total test conducted	28,58,713	11948	314570	58239	1,25,923
Number of detected cases (percentage) of <i>M. tuberculosis</i>	7,79,195 (27.26%)	-	289205 (91.9%)	50311 (86.4%)	11124 (8.83%)
DR-TB detection; number (percentage)	53826 (7.0%) R-resistant	620 (5.2%) XDR-TB	21739 (7.5%) MDR-TB	2920 (5.80%) XDR-TB	340 (3.1%) R-resistant

[Table/Fig-2]: Number of tuberculosis (TB) cases diagnosed by different C&DS based laboratories in 2020-21 [17]. C&DST: Mycobacterial culture and drug sensitivity testing; CBNAAT: Cartridge based nucleic acid amplification test; True NAT: True nucleic acid amplification test; LPA: Line probe assay;

DR-TB: Drug resistance tuberculosis; MDR: Multidrug resistance; XDR: Extensively drug resistance; R-resistant: Rifampicin resistance

Network of Mycobacterium Culture and Drug Sensitivity Test (CDST) Laboratory

Laboratory networks with advanced diagnostic capability determine the efficacy of TB control programme in the new millennium including new technologies have made faster and more reliable of diagnosis, identification, and DST in developing countries like India [47]. The NTEP lab network's especially CDST laboratories are fitted with a variety of diagnostic technology for DR-TB diagnosis, including traditional solid culture and/or newer rapid TB diagnostic technologies, such as the LPA and liquid culture [51]. Existing detection methods include everything from basic smear microscopy and slow culture to advanced, expensive and technically complex molecular assays [23]. NSP (2017-2025) is based on huge network of all the three tier laboratories throughout the country for all the cases including all possible DR-TB [17].

The TB laboratory network has been expanded over the years to provide better access to quality assured diagnostic services [10]. If the analysis of number of laboratories under NTEP are done in these four to five years, we can see the efforts of government towards TB elimination [Table/Fig-3]. According to annual reports issued by India's Ministry of Health, there were 28 CDST laboratories in 2016-2017, but by 2020-21, the number had raised up to 87 laboratories. Same trends are also showing by the CBNAAT and TrueNAT laboratories [Table/Fig-3]. They are radio controlled by National Skilled Committee on identification and management of TB and the apex committee give the technical recommendation for the laboratory policy [12]. Further, NRL coordination committee reviews the progress and facilitates newer initiatives [52].

India has successfully created one of the largest TB laboratory networks in the world with 6 NRL, 31 Intermediate Reference Laboratories, 87 certified laboratories for Liquid Culture and Drug Susceptibility Testing services, and 64 certified laboratories for LPA services along with 21,717 Designated Microscopy Centres. These all laboratories support in the diagnosis of TB and provided patients

S.	Laboratories under NTEP	Total no. of laboratories established year-wise throughout the India						
No.		2016-17	2017-18	2018-19	2019-20	2020-21		
1.	Florescence microscopy	13888	14000	16000	20356	21,717		
2.	CDST	28	37	48	50	87		
3.	NAAT (CBNAAT and TrueNAT)	628	628	1180	1530	3147		
4.	LPA	54	56	62	64	64		
[Table/Fig-3]: Network of different laboratories status as per annual TB reports of India (2017-2021) [10,17,49,51]. NTEP: National tuberculosis elimination programme; C&DST: Mycobacterial culture and drug sensitivity testing; CBNAAT: Cartridge based nucleic acid amplification test; True NAT: True nucleic								

acid amplification test: LPA: Line probe assay

with access to effective treatment depending on their drug resistance patterns. The Indian government has taken several measures to eradicate tuberculosis. At 8000 DMCs, the NTEP programme aims to substitute smear microscopy with upfront molecular testing using NAAT for TB diagnosis. There are currently 3000 NAAT platforms in the NTEP programme, as well as 18 CDST laboratories being built and 28 CDST laboratories being upgraded with LPA [17].

CONCLUSION(S)

To achieve the vision of a TB free India, the NSP proposes ambitious policies with ample funds to completely abolish the TB cases in India by 2025. Three years are only ahead for this nationwide end TB commitments set out in the sustainable development goals. But still there is need of scale-up free, highly sensitive diagnostic tests. Although, there is provision of universal tests for drug-resistant TB (UDR) by the help of CBNAAT and TrueNAT including especial cases of MDR and XDR-TB through LPA. But more numbers of laboratories are still needed to complete the goal of its elimination. Incidence and mortality rates caused by TB have recently declined, but many cases of DR-TB remain undiagnosed or ineffectively handled by unskilled healthcare service providers in India. To tackle this problem Government of India should declare TB as a public health emergency and launch a campaign to fight it. Aside from aggressive TB prevention campaign, stricter and faster diagnostic procedures, as well as continuous or periodic survey of drug resistance, will be preventative measures of chemotherapy. It will also serve as a helpful parameter among previous and current NSP programmes for achieving the abolition of TB in India by 2025.

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REFERENCES

- Peloquin CA, Berning SE. Infection caused by *Mycobacterium tuberculosis*. Annals of Pharmacotherapy. 1994;28(1):72-84.
- [2] Maurya AK, Singh AK, Kumar M, Umrao J, Kant S, Nag VL, et al. Changing patterns and trends of multidrug-resistant tuberculosis at referral centre in Northern India: A 4-year experience. Indian J Med Microbiol. 2013;31(1):40-46.
- [3] World Health Organization 14 Oct 2020: Global Tuberculosis Report; 2019. Available from: https://www.who.int/news-room/fact-sheets/detail/tuberculosis (Last accessed on 2021 March 26).
- [4] Branigan D; October 2020: Pipeline Report; 2020. Available from: https:// www.treatmentactiongroup.org/wp_content/uploads/2020/10/pipeline_TB_ Diagnostics_2020_final.pdf. (Last accessed on 2021 March 24).
- [5] Sharma M, Kumar D, Bohra GK, Meena DS, Bhambu SK. Study of the prevalence of Multidrug-Resistant Pulmonary Tuberculosis (MDR-TB) in Western Rajasthan using line probe assay. J Family Med Prim Care. 2020;28:9(2):1093-97.
- [6] Singhal P, Dixit P, Singh P, Jaiswal I, Singh M, Jain A. A study on pre-XDR & XDR tuberculosis & their prevalent genotypes in clinical isolates of *Mycobacterium tuberculosis* in north India. Indian J Med Res. 2016;143(3):341-47.
- [7] Institute of Medicine (US). Facing the Reality of Drug-Resistant Tuberculosis in India: Challenges and Potential Solutions: Summary of a Joint Workshop by the Institute of Medicine, the Indian National Science Academy, and the Indian Council of Medical Research. Washington (DC): National Academies Press (US); 2012. Available from: http://www.ncbi.nlm.nih.gov/books/NBK100386/. (Last accessed on 2021 April 1).

- [8] Van JI, Simons S, Zwaan R, Laan T, Agterveld M, Boeree M, et al. Comparative study on genotypic and phenotypic second-line drug resistance testing of *Mycobacterium tuberculosis* complex isolates. J Clin Microbiology. 2010;48:2749-53.
- [9] Lohiya A, Suliankatchi AR, Rath RS, Jacob O, Chinnakali P, Goel AD, et al. Prevalence and patterns of drug resistant pulmonary tuberculosis in India-A systematic review and meta-analysis. J Glob Antimicrob Resist. 2020;22:308-16.
- [10] India TB report 2020: Central TB division, Directorate General of Health Services, Ministry of Health and Family Welfare, India. Available from: https://tbcindia.gov. in/showfile.php?lid=3538. (Last accessed on 2021 April 1).
- [11] Dias HMY, Pai M, Raviglione MC. Ending tuberculosis in India: A political challenge & an opportunity. Indian J Med Res. 2018;147(3):217-20.
- [12] National Strategic Plan For Tuberculosis Elimination 2017-2025; March 2017. Central Tb Division, Directorate General of Health Services, Ministry of Health and Family Welfare. Available from: https://tbcindia.gov.in/WriteReadData/ NSP%20Draft%2020.02. 2017%201.pdf (Last accessed on 2021 March 28).
- [13] Singh S, Kumar S. Tuberculosis in India: Road to elimination. Int J Prev Med. 2019;12:10:114.
- [14] Mahadev B, Kumar P. History of tuberculosis control in India. J Indian Med Assoc. 2003;101(3):142-43.
- [15] NTEP-National Tuberculosis Elimination Program: Renamed in 2020. Available from: https://tbfacts.org/rntcp. (Last accessed on 2021 March 26).
- [16] Technical and Operational Guidelines for TB Control in India 2016. New Delhi; Central TB Division, MoHFW. 2016. P.10. Available from: https://tbfacts.org/ rntcp. (Last accessed on 2021 April 02).
- [17] India TB Report 2021; Central TB Division, Directorate General of Health Services, Ministry of Health and Family Welfare. Available from: https://tbcindia. gov.in/showfile.php?lid=3587. (Last accessed on 2021 Feb 25).
- [18] General Laboratory Services in Tuberculosis Control; who/Tb/98.258;WHO, 1998. Available from: https://apps.who.int/iris/bitstream/handle/10665/65942/ WHO_TB_98.258 (part1)/pdf. (Last accessed on 2021 March 26).
- [19] Technical manual for drug susceptibility testing of medicines used in the treatment of tuberculosis; World Health Organisation; 2018. Available from: https://apps. who.int/iris/bitstream/handle/10665/275469/9789241514842-eng.pdf?ua=1. (Last accessed on 2021 March 26).
- [20] Textbook of Microbiology; Ananthnarayan and Panicker seventh edition; 1998. Available from: https://www.moscmm.org/pdf/Ananthanarayan%20microbio.pdf. (Last accessed on 2021 March 26).
- [21] World Health Organization Endorses Truenat Tests for Initial Diagnosis of Tuberculosis and Detection of Rifampicin Resistance; 2 July 2020; ICMR. Available from: https://www.icmr.gov.in/pdf.pressrealease_files/PR_Truenat_ WHO_endorsement_02072020.pdf. (Last accessed on 2021 March 26).
- [22] Gomathi NS, Kumar V. Reliability of Mycobacterial Growth Indicator Tube (MGIT) 960 for the detection of isoniazid resistance in a tuberculosis endemic setting. Indian J Med Res. 2014;139(3):471-73.
- [23] Rakotosamimanana N, Rabodoarivelo MS, Palomino JC, Martin A, Razanamparany VR. Exploring tuberculosis by molecular tests on DNA isolated from smear microscopy slides. Int J Infect Dis. 2017;56:248-52.
- [24] World Health Organization Supra-National Laboratory Network. Available from: http://www.who.int/tb/areas-of-work/laboratory/srl-network/en. (Last accessed on 2021 March 10).
- [25] Parrish NM, Carroll KC. Role of the clinical mycobacteriology laboratory in diagnosis and management of tuberculosis in low-prevalence settings. J Clin Microbiology. 2011;49(3):772-76.
- [26] Acharya R, Vyas M. Comparative study between FM staining and Z-N staining in diagnosing sputum smear positive PTB at S.P. Medical College and Hospital. Int J Med Res Prof. 2018;4(4);134-37.
- [27] Steingart KR, Henry M, Ng V, Hopewell PC, Ramsay A, Cunningham J, et al. Fluorescence versus conventional sputum smear microscopy for tuberculosis: A systematic review. Lancet Infect Dis. 2006;6(9):570-81.
- [28] Manual for Sputum Smear Fluorescence Microscopy; Central TB Division; 1998. Available from: https://tbcindia.gov.in/WriteReadData/892s/7890638455 Flourescence_Microscopy%20Manual.pdf. (Last accessed on 2021 March 12).
- [29] Reza LW, Satyanarayna S, Enarson DA, Kumar AMV, Sagili K, Kumar S, et al. LED-fluorescence microscopy for diagnosis of pulmonary tuberculosis under programmatic conditions in India. PLoS One. 2013;8(10):e75566.
- [30] World Health Organization: Fluorescent Light Emitting Diode (LED) Microscopy for Diagnosis of Tuberculosis; 2010. Available from: https://www.who.int/tb/ laboratory/who_policy_led_microscopy_july10. (Last accessed on 2021 Feb 27).
- [31] Oommen S, Banaji N. Laboratory diagnosis of tuberculosis: Advances in technology and drug susceptibility testing. Indian J Med Microbiol. 2017;35(3):323-31.
- [32] Revised National TB Control Programme Training Manual for Mycobacterium tuberculosis Culture & Drug susceptibility testing; Central TB\Division Directorate General of Health Services Ministry of Health and Family Welfare; April, 2009. Available from: https://tbcindia.gov.in/writeReadData/ I892s/6995271860Training%20manual%20M%20tuberculosis%20C%20DST. pdf. (Last accessed on 2021 March 26).
- [33] Rodrigues C, Shenai S, Sadani M, Sukhadia N, Jani M, Ajbani K, et al. Evaluation of the bactec MGIT 960 TB system for recovery and identification of *Mycobacterium tuberculosis* complex in a high through put tertiary care centre. Indian J Med Microbiol. 2009;27(3):217-21.
- [34] World Health Organization; 26 March 2007. Available from: http://www.who. int/tb/dots/laboratory/policy/en/index3.html. Accessed February 2009. (Last accessed on 2021 March 26).

- [35] Training Modules (1-4) for Programme Managers and Medical Officers; Central TB Division, Ministry of Health and Family Welfare (Mohfw), Government of India; 2020. Available from: https://tbcindia.gov.in/WriteReadData/NTEPTrainingModules1to4.pdf. (Last accessed on 2021 Feb 28).
- [36] Hillemann D, Richter E, Rüsch-Gerdes S. Use of the BACTEC Mycobacteria growth indicator tube 960 automated system for recovery of Mycobacteria from 9,558 extrapulmonary specimens, including urine samples. J Clin Microbiol. 2006;44(11):4014-17.
- [37] Xpert MTB/RIF assay for the diagnosis of pulmonary and extrapulmonary TB in adults and children; World Health Organisation; 2013. Available from: https:// apps.who.int/iris/bitstream/handle/10665/112472/9789241506335_eng. pdf?sequee=1. (Last accessed on 2021 March 26).
- [38] World Health Organisation consolidated guidelines on tuberculosis, Module 4: Treatment drug-resistant tuberculosis treatment; 2020. Available from: https://www. who.int/publications/i/item/9789240007048. (Last accessed on 2021 March 26).
- [39] Sachdeva K, Shrivastava T. CBNAAT: A boon for early diagnosis of tuberculosishead and neck. Indian J Otolaryngol Head Neck Surg, 2018;70(4):572-77.
- [40] Lee DJ, Kumarasamy N, Resch SC, Sivaramakrishnan GN, Mayer KH, Tripathy S, et al. Rapid, point-of-care diagnosis of tuberculosis with novel Truenat assay: Cost effectiveness analysis for India's public sector; PLoS one. 2019;14(7);e0218890.
- [41] Nikam C, Kazi M, Nair C, Jaggannath M, Manoj M, Vinaya R, et. al. Evaluation of the Indian TrueNAT micro RT-PCR device with GeneXpert for case detection of pulmonary tuberculosis. Int J Mycobacteriol. 2014;3(3):205-10.
- [42] Yacoob FL, Philomina Jose B, Karunakaran Lelitha SD, Sreenivasan S. Primary multidrug resistant tuberculosis and utility of line probe assay for its detection in smear-positive sputum samples in a tertiary care hospital in South India. J Pathogens. 2016;2016:6235618.
- [43] Line probe assays for drug-resistant tuberculosis detection interpretation and reporting guide for laboratory staff and clinicians; Global Laboratory Imitative advancing TB diagnosis. Available from: http://www.stoptb.org/wg/gli/assets/ documents/LPA_test_ web_ready.pdf. (Last accessed on 2021 March 20).

- [44] Molecular line-Probe assay for the detection of resistance to isoniazid and rifampicin (LPA); World Health Organisation; 2016. https://www.who.int/tb/ publications/factsheet/tb/ fllpa.pdf. (Last accessed on 2021 March 26).
- [45] Nathavitharana RR, Cudahy PG, Schumacher SG, Steingart KR, Pai M, Denkinger CM. Accuracy of line probe assays for the diagnosis of pulmonary and multidrug-resistant tuberculosis: A systematic review and meta-analysis. Eur Respir J. 2017;18;49(1):1601075.
- [46] Maningi NE, Malinga LA, Antiabong JF, Lekalakala RM, Mbelle NM. Comparison of line probe assay to BACTEC MGIT 960 system for susceptibility testing of first and second-line anti-tuberculosis drugs in a referral laboratory in South Africa. BMC Infect Dis. 2017;17(1):795.
- [47] Azadi D, Motallebirad T, Ghaffari K, Shojaei H. Mycobacteriosis and tuberculosis: Laboratory diagnosis. Open Microbiol Journal. 2018;30;12:41-58.
- [48] Kumar A, Singh AK, Upadhyay V, Pandey J. Epidemiology of multi-drug-resistant tuberculosis in Northern India. Biomed Biotech Res J. 2018;2(2):112-21.
- [49] Annual TB report 2019; Central TB Division, Directorate General of Health Services, Ministry of Health and Family Welfare. Available from: https://tbcindia. gov.in/WriteReadData/India%20TB%20Report%202019.pdf. (Last accessed on 2021 April 02).
- [50] Siddiqi S, Ahmed A, Asif S, Behera D, Javaid M, Jani J, et. al. Direct drug susceptibility testing of *Mycobacterium tuberculosis* for rapid detection of multidrug resistance using the Bactec MGIT 960 system: A multicenter study. J Clin Microbiol. 2012;50(2):435-40.
- [51] INDIA TB REPORT 2018; Central TB Division, Directorate General of Health Services, Ministry of Health and Family Welfare. Available from: https://tbcindia. gov.in/showfile.php?lid=3314. (Last accessed on 2021 March 26).
- [52] RNTCP laboratory network: Overview. Available from: https://tbcindia.gov.in// WriteReadData/l892s/9565488992RNTCP%20laboratory%20network.pdf. (Last accessed on 2021 March 20).

PARTICULARS OF CONTRIBUTORS:

- 1. PhD Scholar, Department of Zoology, Deen Dayal Upadhyaya Gorakhpur University, Gorakhpur, Uttar Pradesh, India.
- 2. Assistant Professor and Head, Department of Microbiology, BRD Medical College, Gorakhpur, Uttar Pradesh, India.
- 3. Assistant Professor, Department of Zoology, Deen Dayal Upadhyaya Gorakhpur University, Gorakhpur, Uttar Pradesh, India.
- 4. Microbiologist, Mycobacterial Culture and Drug Sensitivity Laboratory, Department of Microbiology, BRD Medical College, Gorakhpur, Uttar Pradesh, India.
- Junior Resident, Viral Diagnostic Research Laboratory, Department of Microbiology, BRD Medical College, Gorakhpur, Uttar Pradesh, India.

NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR: Dr. Sushil Kumar.

Assistant Professor, Department of Zoology, Deen Dayal Upadhyaya Gorakhpur University, Gorakhpur, Uttar Pradesh, India. E-mail: sushilk731@gmail.com

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