Microbiology Section

Evaluation of Truenat MTB/RIF Test in Comparison with Microscopy and Culture for Diagnosis of Extrapulmonary Tuberculosis in a Tertiary Care Centre

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ABSTRACT

Introduction: Worldwide, Extrapulmonary Tuberculosis (EPTB) accounts for 15-20% of all cases of TB. The diagnosis of EPTB is a big challenge, as the number of *Mycobacterium tuberculosis* (MTB) bacilli in the tissues and other organs is often very low. Truenat MTB/RIF (rifampicin) is a novel method, which is battery operated, point-of-care and chip-based Real Time Polymerase Chain Reaction (RT-PCR) micro device.

Aim: To evaluate Truenat as a screening test in the diagnosis of EPTB in comparison with microscopy and culture.

Materials and Methods: A prospective cross-sectional study was carried out over a year in which samples from suspected cases of EPTB fitting in the inclusion criteria were subjected to Ziehl-Neelsen (ZN) staining for smear microscopy, culture on Lowenstein Jensen (LJ) medium and PCR for MTB by Truenat. Comparisons were made between the tests and the data was presented using

summary statistics with 95% Confidence Interval (CI).

Results: A total of 248 samples were received from suspected cases of EPTB. Out of the different samples tested, 9 (3.6%) were positive with Truenat MTB. The predominant type of EPTB observed in the study was lymph node Tuberculosis (TB) (66.6%) followed by intestinal, pleural and skeletal TB. Out of the 106 samples tested for culture, four were culture positive for MTB and out of 178 samples tested for microscopy, three were positive for acid fast bacilli. Sensitivity, specificity, Negative Predictive Value (NPV), Positive Predictive Value (PPV), observed agreement of Truenat with culture and microscopy were 100%, 95.1%, 100%, 44.4%, 95.3% and 100%, 96.6%, 100%, 33.3%, 96.6%, respectively.

Conclusion: Truenat MTB test is a cost-effective rapid molecular test with good sensitivity and specificity for the diagnosis of EPTB in low resource settings.

Keywords: Microchip, Mycobacterium tuberculosis, Novel test, Sensitivity, Specificity

INTRODUCTION

Tuberculosis is a major global health problem. Worldwide, TB is one of the top 10 causes of deaths. Globally, the annual incidence of TB estimates about 10.0 million people of which 2.7 million cases are reported from India [1]. As there is no single test that can be used to detect TB in developing countries like India, a novel cost-effective test is required for its diagnosis. The conventional diagnostic modalities have many limitations as it is time consuming, cumbersome and lacks sensitivity. This often delays the early diagnosis and treatment of TB [2].

EPTB affects many organs or sites and is caused by MTB. The various forms of EPTB are lymph node, central nervous system, pleural, abdominal, bone and joint, pericardial, urogenital, cutaneous and ocular. While EPTB accounts for 15-20% of all TB cases in HIV negative patients, its burden in HIV positive patients is high and it is estimated to be 40-50% [3]. Due to the lack of rapid and cost-effective testing methods, there has been a delay in the reduction of TB-related morbidity and mortality. In developing countries, where 95% of new TB cases and death occurs due to TB, smear microscopy remains the most common and often the only test available that detects only 45% of TB infections [4,5].

Smear microscopy by ZN technique is the cornerstone for the diagnosis of TB in resource-limited settings but it has relatively moderate (80%) sensitivity and a poor PPV [6]. Culture is the "gold standard" for final determination, and also permits drug susceptibility testing. However, it takes 2-6 weeks and delays the initiation of treatment [7].

Nucleic Acid Amplification Testing (NAAT) on direct samples is considered to be rapid than culture and have high turn around time as specimens are being sent to distant laboratories. PCR testing is expensive which makes it inaccessible for the patients in TBendemic countries [2]. The commonest target used in PCR test is IS6110 and this has good sensitivity and specificity for the diagnosis of pulmonary and EPTB [8].

The miniaturised forms of PCR tests have the advantages of a reduction in the cost of instruments and faster turn around times in poor resource settings. The micro-PCR devices have the added advantages of better diagnostic sensitivity and portability. They are widely used in India and other South-East Asian countries [9]. GeneXpert was endorsed by the World Health Organisation (WHO) to be used in India as part of the National TB control programme [10]. Cartridge Based Nucleic Acid Amplification Test (CBNAAT) is a test which detects TB bacilli and also screens for rifampicin drug resistance [11]. The assay is optimised and used specifically for case detection of pulmonary TB in the public sector in India. The sensitivity of the assay for EPTB is highly variable, ranging from 25%-96% in different studies [12-15]. Lower sensitivities are reported for samples like Cerebrospinal Fluid (CSF) and other sterile site body fluids [16].

Especially in India, there is a need for a more cost-effective rapid method for diagnosis. Truenat is a novel method, which is a point-of-care, battery operated and chip-based RT-PCR micro device for the detection of MTB in clinical samples. It can be used in low- infrastructure settings to give rapid diagnosis with a very small amount of sample. In Truenat MTB 'detected' samples, the

presence of rifampicin drug resistance can be tested with a second chip as a follow-on test. Truenat MTB test has high sensitivity and specificity for the early diagnosis of pulmonary and EPTB when compared to GeneXpert [17]. However, there is no study where it was validated against culture methods with extrapulmonary specimens. Moreover, since Pathanamthitta district of Kerala has one of the lowest prevalence of TB in the country, the PPV is bound to reduce too. Therefore, it is important to estimate the ability of Truenat MTB to diagnose TB correctly, especially EPTB which has even lower prevalence. The present study was aimed to evaluate Truenat MTB test in comparison with microscopy and culture for the diagnosis of EPTB.

MATERIALS AND METHODS

This was a prospective cross-sectional study which was done in a tertiary care centre in central Kerala, India during the period from January 2019 to December 2019. Total 248 extrapulmonary clinical samples were obtained during this period. This study was conducted after getting clearance from Institutional Ethical Committee (IEC) with registration number ECR/1098/Inst/KL/2018.

Inclusion Criteria: All presumptive EPTB cases irrespective of age, previous history of Anti-Tubercular Treatment (ATT) and whose treatment failed at the end of their most recent course of treatment were included in the study. The different forms of EPTB includes lymph node TB, pleural TB, abdominal TB, central nervous system TB, pericardial TB, spinal TB, bone and joint TB, genitourinary TB, ocular TB and cutaneous TB [18].

Exclusion Criteria: Patients who were on ATT were excluded from the study.

Diagnosis of EPTB: Any clinical sample in which MTB was detected by Truenat MTB test or positive culture/microscopy was considered as a microbiologically proven case of EPTB. Any sample which was positive with culture/microscopy and if MTB was not detected by Truenat was evaluated for presence of Non-Tuberculous Mycobacteria (NTM) by culture [10,19].

Processing of the Clinical Specimens: Various specimens like tissue biopsies, pleural fluid, ascitic fluid, peritoneal fluid, urine, synovial fluid, CSF, bone specimens, liver aspirate and pus from cold abscess/deep sites were obtained from suspected cases after getting informed consent from the patients. All the fluid specimens were centrifuged at 5000 rpm (revolutions per minute) for five minutes in a sterile tube. The tissue specimens are washed with sterile water and they are homogenised by using mortar and pestle or a micro pestle.

Truenat MTB Plus test: Truenat MTB Plus test was performed as per manufacturer's instructions [20,21].

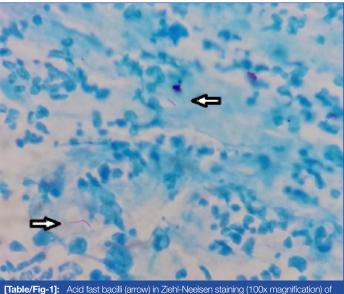
Sample Processing Procedure: All the samples were treated as per Molbio EPTB sample pre-treatment protocol. After discarding the supernatant of the centrifuged specimens, 0.5 mL of the sediment was transferred to Lysis buffer tube. The homogenised tissue sample and pus aspirate were treated with liquefaction buffer for five minutes and then transferred to Lysis buffer tube. The tube was then vortexed for five minutes.

Extraction Procedure of Trueprep Auto kit: The extraction of Deoxyribonucleic Acid (DNA) from the samples was done using Trueprep AUTO Universal Cartridge Based Sample Prep kit and device. The pre-treated sample was transferred to the sample chamber of the cartridge and was placed in the device. The entire elute was aspirated out from the elute chamber into the Elute Collection Tube (ECT).

Truenat MTB Plus Real Time PCR: The 6 μ L of purified DNA from ECT was transferred to microtube containing freeze dried PCR reagents. It was then added to the Truenat MTB microchip containing lyophilised mastermix and the real-time PCR was done using a pre- programmed profile on Truelab Analyser. Results

obtained using the Real-time PCR were reported as 'not detected' for negative samples or 'detected with number of Colony Forming Units per milliliter (CFU/mL)' for positive samples. The Limit of Detection (LoD) in Truenat MTB test was 100 CFU/mL and a follow on test for rifampicin resistance was planned to be performed using Truenat MTB-RIF Dx in those positive samples with more than 10,000 CFU/mL.

Conventional culture and microscopy: The samples after digestion and concentration were processed for culture in Lowenstein Jensen media and for smear microscopy as per standard protocols [22,23]. The culture bottles were examined daily for the initial one week and thereafter weekly once for eight weeks. Ziehl-Neelsen staining was performed and observed for the presence of acid fast bacilli [Table/Fig-1].



[Table/Fig-1]: Acid fast bacilli (arrow) in Ziehl-Neelsen staining (100x magnification) of pus aspirate sample.

Out of 248 samples received, all were subjected to Truenat MTB test. Due to insufficiency of samples and unforeseen reasons, only 107 samples were proceeded for culture and 179 samples for ZN staining. The results of Truenat, microscopy and culture were analysed. Observed percentage of agreement, sensitivity, specificity, PPV and NPV of Truenat were evaluated over culture and microscopy.

STATISTICAL ANALYSIS

Comparisons of sensitivity, specificity, NPV, PPV and observed agreement of Truenat MTB test with other tests were applied. Data was presented using summary statistics with 95% CI. Since this study is first of its kind, the estimation of sample size was overwhelming without prior knowledge of the expected test discordance and may be an underestimate of the sample size required.

RESULTS

A total of 248 suspected cases were studied over a period of one year from January 2019 to December 2019. Samples were obtained from various departments like Surgery, Urology, Medicine, Orthopaedics, Paediatrics, Neurosurgery, Gastroenterology, Pulmonary Medicine and Gynaecology [Table/Fig-2].

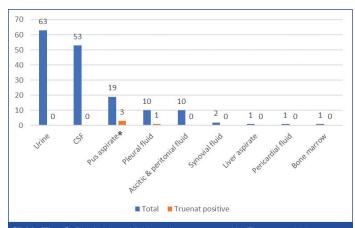
There was an almost equal distribution of males and females among the study population with 126 males (50.8%) and 122 females (49.2%). Maximum number of suspected EPTB patients was in the age group of 50-59 and 60-69 age category (42.3%). Age wise distribution of samples is mentioned in [Table/Fig-3]. A total of 160 (64.5%) fluid samples and 88 (35.5%) tissue sample were received. Out of the different fluid samples, urine (63, 39.4%), CSF (53, 33.1%) and pus aspirate (19, 11.9%) were the majority. Of the tissue samples received, lymph node (36, 40.9%), intestinal biopsy (24, 27.3%) and skeletal (16, 18.2%) constituted

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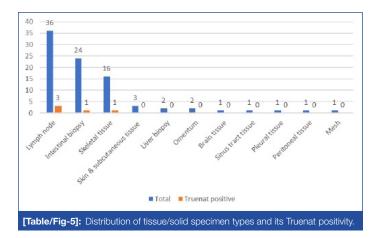
Department	Number of samples (%)				
General Medicine	82 (33.1)				
General Surgery	55 (22.2)				
Urology	52 (21)				
Gastroenterology	26 (10.5)				
Pulmonary Medicine	10 (4)				
Orthopaedics	10 (4)				
Neurosurgery	9 (3.6)				
Paediatrics	3 (1.2)				
Obstetrics and Gynaecology	1 (0.4)				
Total	248				
[Table/Fig-2]: Distribution of samples from different departments.					

Age group (years)	Number of samples	Number of Truenat MTB positive samples	Number of NTM positive samples					
0-9	3							
10-19	14							
20-29	27	3						
30-39	29	2						
40-49	40							
50-59	52	3						
60-69	53		1					
70-79	20	1						
80-89	10							
Total	248	9	1					
[Table/Fig-3]: Age wise distribution of samples.								

MTB: Mycobacterium tuberculosis; NTM: Non tuberculous mycobact







the majority. [Table/Fig-4,5] shows the different fluid and tissue specimens which were studied.

Out of the 248 samples tested, 9 (3.6%) were positive for *MTB* by Truenat. Out of the different positive samples, three were pus aspirates (33.3%), three were lymph node tissue (33.3%) and one each were pleural fluid, intestinal biopsy and skeletal tissue (11.1% each) [Table/Fig-4,5]. All the positive aspirated pus was from lymph node cold abscesses. In the present study, lymph node TB was the predominant type of EPTB (6, 66.6%). Maximum number of positive cases were in the age group category 20-29 (33.3%) and 50-59 (33.3%).

Culture in conventional solid media (LJ) was performed with 106 samples, out of which MTB was isolated in four samples. All culture positive samples were positive for MTB by Truenat also. Apart from these, one sample (infected mesh after herniorrhaphy) which was culture positive for NTM was negative when tested with Truenat MTB. Out of the 179 samples subjected to ZN staining, acid fast bacilli were seen in four samples. Of the smear positive samples, one was later on confirmed with culture as NTM and was excluded from statistical analysis, so sample for ZN was 178.

The effectiveness of Truenat as a screening test for EPTB was assessed by comparing the results with culture as the gold standard and with microscopy. The sensitivity, specificity, PPV, NPV and observed agreement of Truenat with culture and microscopy are summarised in [Table/Fig-6].

		Truenat						Ob-	
Tests	Results	Posi- tive (n)	Neg- ative (n)	Sen- sitiv- ity (%)	Spec- ificity (%)	PPV (%)	NPV (%)	served agree- ment (%)	
Culture (n=106) [‡]	Positive (n)	4	0	100	95.1	44.4	100	95.3	
	Negative (n)	5	97						
Microscopy (n=178) [‡]	Positive (n)	3	0	100	100 96	06.6	33.3	100	96.6
	Negative (n)	6	169			96.6			

[Table/Fig-6]: Comparison of Truenat with culture and microscopy.

*Comparisons were made using summary statistics with 95% Cl. ‡Out of 179 samples for microscopy, one was negative with Truenat which was confirmed as

NTM with culture. Hence, comparison was performed with 178 samples for microscopy and 106 samples for culture.

DISCUSSION

The present study had a high concordance for Truenat against culture and microscopy with high sensitivity, specificity, PPV and NPV showing the effectiveness of this innovative tool from India. But unlike other studies which involved pulmonary samples, this study focused on extrapulmonary samples which is now showing an increasing proportional trend [16,20]. This study is the first of its kind to evaluate the effectiveness of Truenat for extrapulmonary specimens as there is a scarcity of literature on the effectiveness of Truenat in diagnosis of EPTB [24].

This study was conducted with a total of 248 samples of presumptive TB cases received from various clinical departments to the Microbiology and TB unit of a tertiary care teaching institution in central Kerala, India. EPTB was seen more in females (6 out of 9 positives) which are in par with another study [25]. The global reports on TB by WHO found a higher incidence of the disease among males [26]. TB affects mostly the adult age groups in the developing world and this study is no different. Maximum number of positive cases (three each) was in the age group of 20-29 and 50-59 category. Although the highest burden of TB is among adult men, people of all age groups are affected [25]. Lymph node TB was the predominant type in present study (6, 66.6%) followed by intestinal, pleural and skeletal TB (1 each, 11.1%). The most common type of EPTB was lymph node TB followed by pleural TB in other studies [25,27]. Endometrium

was the commonest sample type in a study on the diagnosis of EPTB by Truelab MicroPCR by Ranjana H and Sadhna S [24].

With culture as gold standard test, Truenat had a sensitivity of 100% and specificity of 95.1%. When compared with microscopy, Truenat test had 100% sensitivity and 96.6% specificity. In a similar study conducted by Nikam C et al., for evaluation of Truenat using sputum samples, they found a sensitivity value of 100% and a specificity value of 43.98% for Truenat over microscopy, whereas for Truenat over culture they obtained a sensitivity value of 94.70% and a specificity value of 52.85% [17]. In another study conducted by Nikam C et al., it was reported that the Truenat MTB was able to detect TB rapidly with good sensitivity in comparison with a Composite Reference Standard (CRS) [20]. Present study results were in concordance with this study, though there was higher specificity also. Both the above studies evaluated the effectiveness of Truenat MTB using pulmonary samples unlike present study which is presumed to be the first of its kind in EPTB.

One sample which was Truenat negative was positive in smear microscopy and NTM was isolated in culture. For the diagnosis of PTB and EPTB, culture and microscopy are also of value along with the molecular tests, as Truenat detects only MTB. This was comparable to another study which compared culture, microscopy and GeneXpert [28]. For the detection of both MTB and NTM, multiplex Real Time PCR should be used [29].

The load of bacilli required to obtain a positive value for rifampicin resistance is nearly 10⁴ bacilli however this study was conducted with extrapulmonary specimens where the bacilli load was even less so the authors were unable to perform rifampicin resistance using Truenat in all positive cases. In present study, the bacterial load of positive samples ranged from 100 to 1000. However, in the three positive samples screened, rifampicin resistance was not detected. Rifampicin sensitivity testing in EPTB has to be evaluated in larger sample sizes and the need to depend on alternate methods has to be studied. In a study by Vijayalakshmi J et al., on evaluation of Truenat with pulmonary samples, Rifampicin drug sensitivity was tested with Truenat MTB/RIF micro PCR chip and resistance was detected in 19% of the samples [21].

In the present study, the Truenat MTB test allows detection of TB in approximately 2 to 3 hours and can be utilised in near-care settings to provide quick and accurate diagnosis. Truenat MTB is a good point-of-care, linkage-to-care and treatment test which is cost effective according to Lee DJ et al., study [30]. Truenat MTB test has a higher sensitivity than other conventional diagnostic tests like smear microscopy or culture for MTB. The present study stresses the importance of this new tool, which is indigenous, economical and convenient to use in a low resource setting like India. This study paves and opens windows for larger studies for replacing other molecular diagnostic tests.

Limitation(s)

Authors have compared the effectiveness of Truenat with microscopy and conventional culture for a period of one year. Since the prevalence of EPTB is low, larger studies are required to compare its effectiveness with GeneXpert and MGIT tests to estimate the performance of Truenat MTB. Due to insufficiency of samples, the preference was given to PCR test and not all samples were subjected for microscopy and culture. Rifampicin resistance can be detected only when the bacilli load is more than 10⁴ CFU/ mL. However, most of the samples that was received were with low bacterial load and therefore, rifampicin resistance could not be assessed.

CONCLUSION(S)

Truenat MTB test has high sensitivity and specificity for the case detection of EPTB with a testing time of less than three hours. As

the NPV is high, this will be an ideal test for screening of TB. In the current study, Truenat assay showed a high concordance with other studies on molecular diagnostic tests. Thus, this assay might be a potential, accurate, and rapid method for the detection of EPTB cases in low resource settings.

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REFERENCES

- Global tuberculosis report 2018. Geneva: World Health Organization; 2018. https://apps.who.int/iris/bitstream/handle/10665/274453/9789241565646eng.pdf?ua=1
- [2] Parsons LM, Somoskövi Á, Gutierrez C, Lee E, Paramasivan CN, Abimiku A, et al. Laboratory diagnosis of tuberculosis in resource-poor countries: Challenges and opportunities. Clin Microbiol Rev. 2011;24:314. doi: 10.1128/CMR.00059-10.
- [3] Sharma SK, Ryan H, Khaparde S, Sachdeva KS, Singh AD, Mohan A, et al. Index-TB Guidelines: Guidelines on extrapulmonary tuberculosis for India. Indian J Med Res. 2017;145:448-63. DOI: 10.4103/ijmr.IJMR_1950_16.
- World Health Organization Fact Sheet No. 104: Tuberculosis. Geneva: WHO; 2019. https://www.who.int/en/news-room/fact-sheets/detail/tuberculosis
- [5] Dye C, Watt CJ, Bleed DM, Hosseini SM, Raviglione MC. Evolution of tuberculosis control and prospects for reducing tuberculosis incidence, prevalence, and deaths globally. JAMA. 2005;293(22):2767-75.
- [6] Mathew P, Kuo YH, Vazirani B, Eng RH, Weinstein MP. Are three sputum acidfast bacillus smears necessary for discontinuing tuberculosis isolation? J Clinl Microbiol. 2002;40(9):3482-84.
- [7] Sowjanya DS, Behera G, Reddy VR, Praveen JV. CBNAAT: A novel diagnostic tool for rapid and specific detection of *mycobacterium tuberculosis* in pulmonary samples. Int J of Health Res in Modern Integrated Medical Sciences (IJHRMIMS). 2014;1(1):28-31.
- [8] Cheng VC, Yam WC, Hung IF, Woo PC, Lau SK, Tang BS, et al. Clinical evaluation of the polymerase chain reaction for the rapid diagnosis of tuberculosis. J Clin Pathol. 2004;57(3):281–85. doi: 10.1136/jcp.2003.012658.
- [9] Mathema B, Kurepina NE, Bifani PJ, Kreiswirth BN. Molecular epidemiology of tuberculosis: Current insights. ClinI Microbiol Rev. 2006;19(4):658-85.
- [10] Xpert MTB/RIF implementation manual: technical and operational 'howto'; practical considerations. World Health Organization; 2014. https:// apps.who.int/iris/bitstream/handle/10665/112469/9789241506700_eng. pdf?sequence=1&isAllowed=y
- [11] Kasat S, Biradar M, Deshmukh A, Jadhav S, Deshmukh H. Effectiveness of CBNAAT in the diagnosis of extrapulmonary tuberculosis. Int J Res Med Sci. 2018;6(12):3925-28.
- [12] Lawn SD, Mwaba P, Bates M, Piatek A, Alexander H, Marais BJ, et al. Advances in tuberculosis diagnostics: The Xpert MTB/RIF assay and future prospects for a point-of-care test. The Lancet Infect Dis. 2013;13(4):349–61.
- [13] Causse M, Ruiz P, Juan Bautista GA, Casal M. Comparison of two molecular methods for the rapid diagnosis of extrapulmonary tuberculosis. J Clin Microbiol. 2011;49:3065–67.
- [14] Hillemann D, Rusch-Gerdes S, Boehme C, Richter E. Rapid molecular detection of extrapulmonary tuberculosis by the automated GeneXpert MTB/RIF system. J Clin Microbiol. 2011;49:1202–05.
- [15] Vadwai V, Boehme C, Nabeta P, Shetty A, Alland D, Rodrigues C. Xpert MTB/RIF, A new pillar in the diagnosis of extrapulmonary tuberculosis? J Clin Microbiol. 2011;49:2540–45.
- [16] Friedrich SO, von Groote-Bidlingmaier F, Diacon AH. Xpert MTB/RIF assay for diagnosis of pleural tuberculosis. J Clin Microbiol. 2011;49(12):4341-42.
- [17] Nikam C, Kazi M, Nair C, Jaggannath M, Manoj M, Vinaya R, et al. Evaluation of the Indian TrueNAT micro RT-PCR device with GeneXpert for case detection of pulmonary tuberculosis. Int J Mycobacteriol. 2014;3(3):205-10.
- [18] Index-TB guidelines. Guidelines on extra-pulmonary tuberculosis for India. WHO; 2016. https://apps.who.int/iris/bitstream/handle/10665/278953/IND-tb-guidelines-eng.pdf?sequence=5&isAllowed=y
- [19] Rice JP, Seifert M, Moser KS, Rodwell TC. Performance of the Xpert MTB/ RIF assay for the diagnosis of pulmonary tuberculosis and rifampin resistance in a low-incidence, high-resource setting. PLoS One. 2017;12(10):e0186139. https://doi.org/10.1371/journal.pone.0186139
- [20] Nikam C, Jagannath M, Narayanan MM, Ramanabhiraman V, Kazi M, Shetty A, et al. Rapid diagnosis of *Mycobacterium tuberculosis* with Truenat MTB: A nearcare approach. PLoS One. 2013;8(1):e51121.
- [21] Vijayalakshmi J, Surekha A, Renuka devi A, Uma devi S. Truenat A novel diagnostic tool for rapid detection of *Mycobacterium tuberculosis* and rifampicin resistance in pulmonary samples. Int J Curr Microbiol App Sci. 2019;8(10):1-9.
- [22] Laboratory services in tuberculosis control. Part II. Microscopy. WHO/ TB/98.258. Geneva, Switzerland: WHO, 1998. https://www.who.int/docstore/ gtb/publications/whodoc/who_tb-98-258/en/98.258_microscopy-2.pdf
- [23] Mycobacteriology Laboratory Manual. Global Laboratory Initiative of Stop TB Partership. 2014;1. https://www.who.int/tb/laboratory/mycobacteriologylaboratory-manual.pdf

- [24] Ranjana H, Sadhna S. Truelab MicroPCR in diagnosis of extrapulmonary tuberculosis-our experience. J Microbiol and Related Res. 2016;2:119-26.
- [25] Gaur PS, Suryakant, Bhaskar R, Singh S, Saxena P, Agnihotri S. Incidence and clinical profiles of pulmonary and extra-pulmonary tuberculosis patients in north Indian population: A hospital based retrospective study. Int J Res Dev Pharm L Sci. 2017;6(5):2773-78. Doi: 10.13040/IJRDPL.2278-0238.6 (5).2773-2778.
- [26] Global tuberculosis report 2019. Geneva: World Health Organization; 2019. https:// apps.who.int/iris/bitstream/handle/10665/329368/9789241565714-eng.pdf
- [27] Prakasha SR, Suresh G, D'sa IP, Shetty SS, Kumar SG. Mapping the pattern and trends of extrapulmonary tuberculosis. J Glob Infect Dis. 2013;5(2):54-59.

doi:10.4103/0974-777X.112277

- [28] Arora D, Dhanashree B. Utility of smear microscopy and GeneXpert for the detection of *Mycobacterium tuberculosis* in clinical samples. GERMS 2020;10(2):81-87. doi: 10.18683/germs.2020.1188
- [29] Negi SS, Singh P, Chandrakar S, Gaikwad U, Das P, Bhargava A, et al. Diagnostic evaluation of multiplex real time PCR, GeneXpert MTB/RIF assay and conventional methods in extrapulmonary tuberculosis. J Clin Diag Res. 2019;13(1):12-16.
- [30] Lee DJ, Kumarasamy N, Resch SC, Sivaramakrishnan GN, Mayer KH, Tripathy S, et al. Rapid, point-of-care diagnosis of tuberculosis with novel Truenat assay: Cost-effectiveness analysis for India's public sector. PLoS ONE. 2019;14(7):e0218890. ttps://doi.org/10.1371/journal.pone.0218890

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