

# Aetiological Profile and Antibiotic Susceptibility Pattern in Patients with Urinary Tract Infection in Tripura

JAYANTA DEBNATH<sup>1</sup>, PRADIP KR. DAS<sup>2</sup>, MUNMUN DEBNATH<sup>3</sup>, K.K. HALDAR<sup>4</sup>**Keywords:** Antibiotic susceptibility, Empirical, Urinary tract infection

Though urinary tract infection (UTI) stands as the second most common infection in the community [1]. Treatment of community acquired UTI is usually empirical in resource limited health care settings or while awaiting for the antimicrobial susceptibility test report. To formulate an empirical treatment guideline, appropriate knowledge of local and national antimicrobial resistance and aetiological trends is of utmost importance [2]. Also, considering the fact, that resistance rates to chemotherapeutics commonly used in UTI is increasing and susceptibility of micro-organisms shows significant geographical variations, recent studies to enhance knowledge on changing trends of aetiological agents in UTI and their antibiotic susceptibility pattern at local levels, serve an important guide for empirical treatment [3].

The present study was conducted for a period of two years, from June 2011 to May 2013, at Dr.B.R. Ambedkar Memorial Teaching Hospital, a tertiary health care centre in the state of Tripura. As, no previous reference was available from this state, prevalence of 50% and Confidence interval as 95%, has been assumed for the minimum sample size estimation.

Freshly voided mid stream urine specimens were collected from patients with clinical suspicion of UTI. The samples were cultured for isolation and identification of the pathogenic bacteria. Bacterial counts of more than 105 CfU/ml were considered significant and bacterial counts of 104-105 CfU/ml with significant history of UTI, morbidity and immunocompromised state were followed up and reported as UTI. Antibiotic susceptibility pattern was determined

by Kirby Bauer disc diffusion method as per CLSI guidelines [4]. Extended Spectrum Beta Lactamases (ESBL) were detected for *E.coli* and *Klebsiella spp.* by double disc synergy test [4]. Out of 2396 urine samples processed, 1084 (45.2%) yielded significant bacterial isolates. The predominant isolate was *Escherichia coli* (43.9%) followed by *Enterococcus faecalis* (21.4%), *Klebsiella pneumoniae* (11.8%), *Staphylococcus aureus* (10.3%) and others. Females were more affected than males (3.1: 1), with majority of infections in the age group of 31-45 years (36%) followed by 16-30 years (29.3%).

The Gram negative isolates were highly sensitive to Imipenem (100%), followed by Piperacillin tazobactam (95.9%), Nitrofurantoin (87.3%) and Amikacin (83.2%). Cotrimoxazole (5.8%) was the least sensitive antibiotic for Gram negative isolates followed by Amoxycillin-clavulanic acid (49.1%) and Ciprofloxacin (50.9%). The predominant isolate *E.coli*, exhibited the similar pattern of susceptibility. Gram positive isolates were more sensitive to Vancomycin (95.9%), Amikacin (89.8%), Amoxycillin-clavulanic acid (84.7%), Nitrofurantoin (83.7%) and Gentamicin (80.6%). Ampicillin (25.5%) and Erythromycin (49%) were least sensitive antibiotics. *Enterococci spp.* as the predominant isolate, was highly sensitive to Vancomycin (93.1%) followed by Amikacin and Amoxycillin-clavulanic acid. Extended Spectrum Beta Lactamases (ESBL) was detected in *E.coli* (10.3%) and *Klebsiella pneumoniae* (3.3%). The result of Antibiotic susceptibility pattern has been depicted in [Table/ Fig-1,2].

Bacterial isolate	Total No.	No. of isolates sensitive to Antimicrobial agents - n (%)											
		IMP	PIT	AK	NT	OF	CIP	LEV	AMC	CXM	CTX	CFT	COT
<i>E. coli</i>	476	476 (100)	456 (95.9)	396 (82.7)	420 (87.3)	328 (68.9)	252 (52.9)	380 (79.8)	252 (52.9)	348 (73.1)	332 (69.7)	364 (76.5)	28 (5.9)
<i>Klebsiella spp.</i>	128	128 (100)	120 (93.8)	112 (87.5)	112 (87.5)	92 (71.9)	60 (46.8)	96 (75)	56 (43.8)	84 (65.6)	80 (62.5)	92 (71.9)	0 (0)
<i>Citrobacter spp.</i>	24	24 (100)	24 (100)	24 (100)	24 (100)	20 (83.3)	20 (83.3)	24 (100)	16 (66.7)	24 (100)	20 (83.3)	20 (83.3)	8 (33.3)
<i>Enterobacter spp.</i>	24	24 (100)	24 (100)	20 (83.3)	24 (100)	20 (83.3)	16 (66.7)	24 (100)	16 (66.7)	20 (83.3)	16 (66.7)	20 (83.3)	4 (16.7)
<i>Acinetobacter spp.</i>	20	20 (100)	20 (100)	12 (60)	12 (60)	8 (40)	4 (20)	12 (60)	0 (0)	4 (20)	0 (0)	4 (0)	0 (0)
<i>Pseudomonas spp.</i>	12	12 (100)	12 (100)	4 (33.3)	4 (33.3)	0 (0)	0 (0)	4 (33.3)	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)
<i>Proteus spp.</i>	8	8 (100)	8 (100)	8 (100)	8 (100)	4 (50)	0 (0)	8 (100)	0 (0)	4 (50)	4 (50)	8 (100)	0 (0)
TOTAL (Sensitive)	692	692 (100)	664 (95.9)	576 (83.2)	604 (87.3)	472 (68.2)	352 (50.9)	548 (79.2)	340 (49.1)	484 (69.9)	452 (65.3)	508 (73.4)	40 (5.8)

**[Table/Fig-1]:** Antibiotic sensitivity pattern of Gram negative bacteria isolated from urine culture

IMP – Imipenem ; PIT – Piperacillin Tazobactam ; AK – Amikacin ; NT – Nitrofurantoin ; OF – Ofloxacin ; CIP – Ciprofloxacin ; LEV – Levofloxacin ; AMC – Amoxycillin Clavulanic acid ; CXM – Cefuroxime ; CTX – Ceftriaxone ; CFT – Cefotaxime ; COT – Cotrimoxazole

Bacterial isolate	Total No.	No. of isolates sensitive to Antimicrobial agents - n (%)										
		VA	GEN	AK	NT	OF	CIP	AMP	AMC	CXM	CEP	ER
<i>Enterococci spp.</i>	232	216 (93.1)	168 (72.4)	208 (89.7)	184 (79.3)	176 (75.9)	136 (58.6)	76 (32.8)	188 (81)	172 (74.1)	124 (53.4)	100 (43.1)
<i>Staphylococcus aureus</i>	112	112 (100)	100 (89.3)	96 (85.7)	96m (85.7)	84 (75)	64 (57.1)	12 (10.7)	96 (85.7)	88 (78.6)	76 (67.9)	64 (57.1)
<i>Staphylococcus saprophyticus</i>	48	48	48 (100)	48 (100)	48 (100)	44 (91.7)	36 (75)	12 (25)	48 (100)	40 (83.3)	32 (66.7)	28 (58.3)
TOTAL (Sensitive)	392	376 (95.9)	316 (80.6)	352 (89.8)	328 (83.7)	304 (77.6)	236 (60.2)	100 (25.5)	332 (84.7)	300 (76.5)	232 (59.2)	19 (49)

**[Table/Fig-2]:** Antibiotic sensitivity pattern of Gram positive bacteria isolated from urine culture

VA – Vancomycin ; GEN – Gentamicin ; AK – Amikacin ; NT – Nitrofurantoin ; OF – Ofloxacin ; CIP – Ciprofloxacin ; AMP – Ampicillin ; AMC – Amoxycillin Clavulanic acid ; CXM – Cefuroxime ; CEP – Cephalexin ; ER – Erythromycin

The positivity rate reported in this study was higher than values reported in India 10.86% [5] and 9.17% [6]. This indicates that urine culture was essential for definitive diagnosis of UTI. The age group exhibiting the highest occurrence of UTI was 16-45y. This was the most sexually active group and women of child bearing age fall in this group. The propensity of young women to develop UTI has been explained on the basis of their anatomy (short urethra) and behavioral factors [7].

The predominant isolate was *Escherichia coli* (43.9%) followed by *Enterococcus faecalis* (21.4%), irrespective of the gender. This was unusual because, in most of the previous studies *E.coli* and *Klebsiella spp.* were found to be major pathogens, with a low isolation rate of *Enterococcus spp.* In 1997 and 1998, *Enterococcus faecalis* was isolated in only 0.8% and 0.5% cases of UTI [8]. In 2001, a study reported isolation rate of 8.9% from Mumbai and in 2002, another study from Chandigarh reported isolation rate of *Enterococcus spp.* as 6.4% [9,10]. A recent study from Bihar, reported isolation rate of *Enterococcus spp.* as high as 15.64% [11], which can be compared with the present study. This indicates the emergence of *Enterococcus spp.* as a significant pathogen.

ISDA guidelines recommend a bench mark of 10-20% resistance at which first line empiric therapy should be modified [12]. Considering the susceptibility pattern of the predominant isolates, Nitrofurantoin and Amikacin can be recommended as first line empirical therapy by oral and parenteral route respectively. As Nitrofurantoin is a urinary antibiotic and not frequently used in this part of India, a low level of resistance was observed. Imipenem, Piperacillin-tazobactam and Vancomycin showed least levels of resistance to all organisms, but these antibiotics should be reserved as final therapeutic options, thereby reducing the antibiotic pressure and abuse. Our observation may be correlated with a study from Mangalore, recommending Nitrofurantoin and Amikacin as drugs of choice for treatment of UTI [1]. Considerable resistance of organisms to Ciprofloxacin, Ofloxacin and Levofloxacin were observed in the present study, for which fluoroquinolones can no longer be advocated for empirical therapy. Most of the ESBL producing strains were multi-drug resistant and

showed susceptibility to Imipenem and Piperacillin-tazobactam only.

## CONCLUSION

In this geographical area of North East India, the present data represents *E.coli* as the most predominant organism, followed by *Enterococcus faecalis*, as an emerging pathogen causing UTI. As usual, females were more infected than males, with predominance of infection in reproductive age group. Laboratory diagnosis is essential for definitive diagnosis of UTI. On analysis of local antibiotic susceptibility pattern, Nitrofurantoin and Amikacin represents the choice of first line empiric therapy for treatment of UTI. Due to variation in such scenario in geographical areas and changing pattern of antibiotic susceptibility, more studies need to be conducted at intervals, based on which hospital policies are to be modified.

## REFERENCES

- Pai V, Nair B. Aetiology and sensitivity of uropathogens in outpatients and inpatients with urinary tract infection: Implications on empiric therapy. *Ann Trop Med Public Health*. 2012;5:181-84.
- Akoachere JF, Yvonne S, Akum NH, Seraphine EN. Aetiological profile and antimicrobial susceptibility of community-acquired urinary tract infection in two Cameroonian towns. *BMC Res Notes*. 2012;5:219.
- Gupta K, Scholes D, Stamm WE. Increasing prevalence of antimicrobial resistance causing acute uncomplicated cystitis in women. *J Am Med Assoc*. 1999; 281:736-38.
- Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing; 16th informational supplement. M100-S16. Clinical and Laboratory Standards Institute, Wayne, PA. 2006.
- Akram M, Shahid M, Khan A. Etiology and antibiotic resistance patterns of community acquired urinary tract infections in JNMC Hospital Aligarh, India. *Ann Clin Microb Antimicrob*. 2007; 6:4.
- Eshwarappa M, Dosegowda R, Aprameya IV, Khan MW, Kumar PS, Kempegowda P. Clinico-microbiological profile of Urinary tract Infection in South India. *Indian J Nephrol*. 2011; 21(1):30-36.
- Manges AR, Natarajan P, Solberg OD, Dietrich PS, Riley LW. The changing prevalence of drug resistant *Escherichia coli* clonal groups in a community: evidence for community outbreaks of urinary tract infections. *Epidemiol Infect*. 2006;134:425-32.
- Ram S, Gupta R, Gaheer M. Emerging antibiotic resistance among uropathogens. *Indian J Med Sci*. 2000; 54(9):388-94.
- Desai PJ, Pandit D, Mathur M, Gogate A.. Prevalence, identification and distribution of various species of Enterococci isolated from clinical specimen with special reference to urinary tract infection in catheterized patients. *Indian J Med Microbiol*. 2001;19(3):132-37.
- Gupta V, Yadav A, Joshi RM. Antibiotic resistance pattern in uropathogens. *Indian J Med Microbiol*. 2002; 20(2):96-98.
- Dhananjay K, Sangeeta D, Krishnan N, Aninda S, Udayan G. Current status of uropathogens with special reference to Enterococcus. *Journal of Evolution of Medical and Dental Sciences*. 2013; 2(19) : 3494-504.
- Warren JW, Abrutyn E, Hebel JR, Schaeffer AJ, Stamm WE. Guidelines for antimicrobial treatment of uncomplicated acute bacterial cystitis and acute pyelonephritis in women. *Clin Infect Dis*. 1999, 29:745-59.

### PARTICULARS OF CONTRIBUTORS:

- Assistant Professor, Department of Microbiology, Tripura Medical College & Dr. B.R. Ambedkar Teaching Hospital, Hapania, Agartala, Tripura (West), India.
- Associate Professor, Department of Microbiology, Tripura Medical College & Dr. B.R. Ambedkar Teaching Hospital, Hapania, Agartala, Tripura (West), India.
- Tutor, Department of Microbiology, Tripura Medical College & Dr. B.R. Ambedkar Teaching Hospital, Hapania, Agartala, Tripura (West), India.
- Professor, Department of Microbiology, Tripura Medical College, Agartala, Tripura (West), India.

### NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Dr. Jayanta Debnath,  
Assistant Professor, Department, of Microbiology, Tripura Medical College & Dr. B.R. Ambedkar Teaching Hospital,  
Hapania, Agartala- 799014, Tripura (West), India.  
Phone : 09436120144, Email : drjdebath@gmail.com

FINANCIAL OR OTHER COMPETING INTERESTS: None.

Date of Submission: Jan 29, 2014  
Date of Peer Review: Apr 24, 2014  
Date of Acceptance: May 19, 2014  
Date of Publishing: Aug 20, 2014