

Oxidative Stress in the Development of Genetic Generalised Epilepsy: An Observational Study in Southern Indian Population

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ABSTRACT

Introduction: Oxidative stress resulting from excessive generation of Reactive Oxygen Species (ROS) plays a significant role in neurodegeneration associated with seizures/epilepsy.

Aim: To evaluate oxidative stress markers and antioxidant enzymes in Genetic Generalised Epilepsy (GGE) and to know the extent of oxidative stress induced by Anti-Epileptic Drugs (AEDs) with the time duration of treatment.

Materials and Methods: In this case-control study, 310 GGE patients (male:female=203:107), who were on AED treatment (n=235) and 75 untreated patients (male:female=49:26) along with 310 age and sex matched healthy controls were recruited. Oxidative stress markers such as Nitric Oxide (NO), Malondialdehyde (MDA) and antioxidant enzyme activities namely Superoxide Dismutase (SOD), Glutathione Peroxidase (GPx) and Catalase (CAT) were measured spectrophotometrically.

Results: Significantly higher levels of serum NO, MDA and low levels of plasma Total Antioxidant Capacity (TAC) were

found in patients as compared to controls ($p < 0.001$) whereas erythrocyte SOD, CAT and GPx activities were found to be significantly low in patients when compared to the control group ($p < 0.001$). Statistically significant higher levels of NO, MDA and lower levels of SOD, CAT and TAC were observed in patients subgroup, who were on AEDs for more than >5 years compared to other groups (≤ 1 year and $1 \leq 5$ years) ($p = 0.02$, $p = 0.01$, $p = 0.001$, $p = 0.01$ and $p = 0.05$ respectively). Further, significant increase in the levels of NO, MDA and decreased activities of SOD, CAT were found in treated patients compared to untreated patients ($p < 0.05$) denoting that additional oxidative stress induced by AEDs which results in seizure recurrence and drug intractability.

Conclusion: Our study demonstrated that GGE patients have additional oxidative stress due to AEDs and decreased antioxidant enzyme activities causing an imbalance between oxidant and antioxidant status, which might contribute to the pathogenesis of GGE.

Keywords: Antioxidant enzymes, Antiepileptic drugs, Intractable epilepsy, Reactive oxygen species, Seizure

INTRODUCTION

Epilepsy is a heterogeneous neurological disorder affecting significant proportion of the population (~4%) at some time in life [1]. GGE (erstwhile known as Idiopathic Generalized Epilepsy (IGE)) accounts for one-third of all epilepsy cases [2], characterised by alone or combination of generalized seizures (absence, myoclonus and tonic-clonic seizures) with Electroencephalogram (EEG) hallmark of paroxysms of generalized spike-wave. Oxidative stress, defined as a persistent imbalance between the production of ROS and antioxidant defences [3], often results in irreversible cellular damage. Oxidative stress resulting from excessive generation of ROS has been implicated in the initiation as well as the progression of epilepsy [4] and also in the pathogenesis of many other neurodegenerative disorders including Alzheimer's disease and Parkinson's disease [5]. The ROS generated during normal cellular metabolism can be scavenged by enzymatic antioxidants namely SOD, GPx, CAT, or non-enzymatic antioxidants mainly vitamin E and reduced form of Glutathione (GSH) antioxidant defence mechanisms. However, oxidative stress results either due to increased ROS production or decreased antioxidant systems or both [6].

Accumulating evidence suggests that the chronic usage of AEDs such as valproic acid, phenytoin and carbamazepine may also increase free radical generation followed by oxidative damage in neurons [7].

Lipid peroxidation has been associated with pathological mechanisms involved in various neurological and neurodegenerative diseases [8]. Neurons are more predisposed to damaging effect of oxidative

stress, due to the high concentration of Polyunsaturated Fatty Acid (PUFA) that is susceptible to lipid peroxidation [9].

NO is a signaling molecule in the Central Nervous System (CNS) and reacts with excess ROS to generate Reactive Nitrogen Species (RNS) like peroxynitrite, which is a powerful oxidizing agent that can cause lipid peroxidation, [10] thus contributing to free radical-mediated damage.

GPx is an enzyme consisting of selenocysteine residue, catalyses H_2O_2 reduction by using GSH which is present in cells in high concentrations and thus protecting cellular environment. CAT protects the cells against H_2O_2 by catalysing the reduction of H_2O_2 . However, it has a negligible role at low levels of hydrogen peroxide but becomes more important at higher concentrations of H_2O_2 [11].

Although the role of free radicals has gained importance in neurochemical research, very few studies have been published on the role of antioxidants in GGE. Further, the relation between the antioxidant enzymes and different GGE types has not been reported. Hence, the present study was designed with an aim to evaluate oxidative stress markers, antioxidant enzymes in GGE patients, and to know the extent of oxidative stress induced by AEDs with the time duration of treatment.

MATERIALS AND METHODS

Three hundred and ten, GGE diagnosed patients (male: female=203:107), who were on AED treatment (n=235), and 75 untreated patients (male:female=49:26) along with 310 age and sex matched healthy controls were recruited for the present study

from Nizam's Institute of Medical Sciences, Hyderabad, India, from October, 2012 to December, 2014 with their written informed consent. The present study was approved by Ethical Committee of the "Institute of Genetics and Hospital for Genetic Diseases" as well as the study hospital. The clinical diagnosis was made according to admitted criteria by International League Against Epilepsy (ILAE) [12] on the basis of age of onset, type of seizures and EEG findings. In addition, patient's Magnetic Resonance Imaging (MRI) of the brain was normal. The demographic characteristics including age, gender, age of onset of the disease, duration of the disease, and family history were collected using a structured questionnaire. The information about seizure type, its duration, frequency and AED medical history was also collected. Patients having diabetes mellitus, hypertension, acute, chronic renal failure and other metabolic disorders that are known to cause oxidative stress, were excluded from the study. Also, patients with smoking, tobacco chewing and alcohol habit were also excluded.

As IGE has an age related onset, we have divided the all IGE patients into "classical" IGE, with an onset before 18 years of age and adult onset IGE, epilepsy of later onset i.e., after 18 years of age on the basis of published reports [13] and the concept in the International Classification [12]. Again the patients were categorised into three groups depending on the time duration of AEDs (phenytoin, carbamazepine, sodium valproate etc.) treatment (≤ 1 year, 1- ≤ 5 years and >5 years).

About, 8 ml of venous blood was collected in plain sterile tubes from all the subjects, in case of patients blood was collected within 24 hours from the onset of seizure attack. A part of it was transferred to sterile Ethylenediaminetetraacetic Acid (EDTA) coated vacutainers for the estimation of erythrocyte SOD, GPx, CAT and plasma TAC. Remaining sample was transferred to sterile silicon tubes without addition of anticoagulants and centrifuged at 2000 rpm for 20 minutes to estimate, NO and MDA. Serum NO levels (nitrite and nitrate) (mmol/mL) were determined according to the method based on the diazotisation of sulfanilic acid by NO at acid pH and subsequent coupling to N-(1-naphthyl-ethylenediamine) [14]. MDA levels (nmol/mL) were measured by Buege JA and Aust SD, method using Thiobarbituric Acid (TBA) [15] and TAC by Ferric Reducing Ability of Plasma (FRAP) assay [16]. The erythrocyte GPx activity was estimated by Rotruck JT et al., method [17], SOD by Marklund SL and Marklund G method [18], CAT by Aebi H method [19].

STATISTICAL ANALYSIS

The difference in the levels of oxidative stress markers and activities of antioxidant enzymes between patients and controls was evaluated by Student's t-test whereas, significance among patient subgroups who were on AEDs (such as phenytoin, carbamazepine, sodium valproate etc.) treatment for different time periods (≤ 1 year, 1- ≤ 5 years and >5 years) were analysed by one-way Analysis of Variance (ANOVA) and the p-value of <0.05 was considered to be significant.

RESULTS

Out of 310 patients diagnosed with GGE, 195 (62.9%) patients indicating the majority had the onset before the age of 18 years (classical IGE) whereas, remaining 115 (37.1%) cases had the onset after 18 years of age (adult onset IGE). There was a higher proportion of females, 101(51.8%) in <18 years group (classical IGE), and similarly males, 64 (55.7%) in >18 years group (adult onset IGE). There was no significant difference in the proportions with a family history of epilepsy between "classical" group, 40 (20.5%) and "adult onset" group, 29 (25.2%). In both the age groups, the tonic-clonic variety of seizures and epilepsy types were more common. Similarly, only TC seizures epilepsy type was predominantly observed in 95 patients (48.7%) and 89 patients (77.4%) respectively in both "classical" as well as adult onset IGE groups [Table/Fig-1]. Division

of patients in both classical and adult onset IGE was done [Table/Fig-2]. On the basis of seizure type, the patients with adult onset IGE were divided into three groups. Those with absence of tonic-clonic seizures were defined as "adult onset absence epilepsy"; the combination of myoclonic jerks and tonic-clonic seizures was defined as "adult onset myoclonic epilepsy" and patients with only tonic-clonic seizures were classified as "adult onset tonic-clonic epilepsy" [20].

The levels of serum NO, MDA, plasma TAC, activities of erythrocyte SOD, GPx and CAT in IGE patients and controls have been given in [Table/Fig-3].

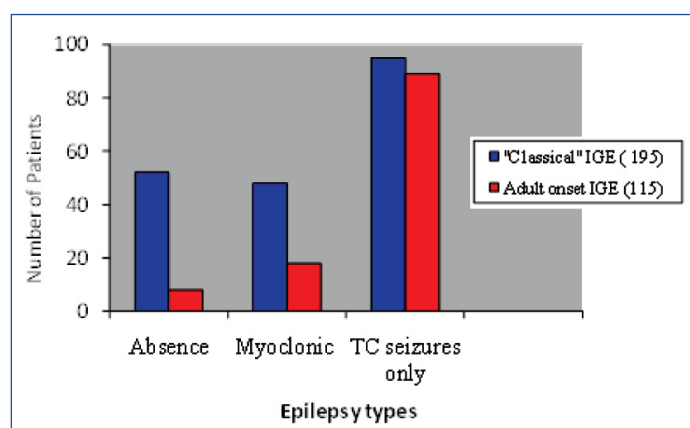
Significantly higher levels of serum NO, MDA and low levels of plasma TAC were observed in patients (8.55 ± 2.0 $\mu\text{mol/mL}$; 4.39 ± 1.65 nmol/mL and 5.47 ± 1.23 $\mu\text{mol/L}$) as compared to controls (5.03 ± 1.27 , 2.42 ± 0.87 and 8.58 ± 1.36) ($p < 0.001$) whereas activities of SOD, CAT and GPx were found to be significantly low in patients when compared to the control group (606 ± 88 U/g Hb, 5.7 ± 1.24 k/mL,

Variables	Age of onset <18 years (n = 195)	Age of onset >18 years (n = 115)
Sex		
Male	94 (48.2)	64 (55.7)
Female	101 (51.8)	51 (44.3)
Family History	40 (20.5)	29 (25.2)
Seizure types		
Absence	71 (36.4)	3 (2.6)
Myoclonus	36 (17.9)	92 (80)
Tonic clonic	181 (92.8)	112 (97.3)
Epilepsy types		
Absence	52 (26.7)	8 (6.9)
Myoclonus	48 (24.6)	18 (15.7)
Tonic clonic	95 (48.7)	89 (77.4)

[Table/Fig-1]: Comparison of characteristics of patients with "classical" and adult onset idiopathic generalized epilepsy.

Note: Values are n (%) unless stated.

On the basis of predominant seizure type and age of onset, the international classification recognizes four IGE sub-syndromes [12] viz., Absence-Absence epilepsy, includes cases of both childhood absence epilepsy (CAE) and juvenile absence epilepsy (JAE); Myoclonic- Myoclonic epilepsy, includes juvenile myoclonic epilepsy (JME); TC- Tonic-clonic seizures, includes idiopathic generalized epilepsy and generalized tonic-clonic seizures on awakening (IGE+GTCS on awakening)



[Table/Fig-2]: Different epilepsy types of "classical" and adult onset IGE with number of patients.

Parameters	Controls (n = 310)	Patients (n = 310)	p-value
NO ($\mu\text{mol/mL}$)	5.03 ± 1.27	8.55 ± 2.0	$<0.0001^*$
MDA (nmol/mL)	2.42 ± 0.87	4.39 ± 1.65	$<0.0001^*$
SOD (U/gHb)	843 ± 95.2	606 ± 88	$<0.0001^*$
CAT (k/ml)	9.39 ± 1.33	5.7 ± 1.24	$<0.0001^*$
GPx (U/gHb)	76.7 ± 11.7	53.8 ± 8.85	$<0.0001^*$
TAC ($\mu\text{mol/L}$)	8.58 ± 1.36	5.47 ± 1.23	$<0.0001^*$

[Table/Fig-3]: The serum MDA, NO, plasma TAC and erythrocyte antioxidant enzyme activities in GGE patients and controls.

Note: p-value $<0.05^*$ is significant

Parameters	0=1 year (n = 54)	1 = 5 years (n = 75)	>5 years (n = 106)	p-value
NO (µmol/mL)	8.1 ± 1.59	8.42 ± 2.02	8.98 ± 2.14	0.02*
MDA (nmol/mL)	4.1 ± 1.34	4.39 ± 1.68	4.87 ± 1.77	0.01*
SOD (U/gHb)	622 ± 92.6	606 ± 88.7	573 ± 67.3	0.001*
CAT (k/ml)	6.11 ± 1.16	5.69 ± 1.26	5.49 ± 1.22	0.01*
GPx (U/gHb)	54.4 ± 8.68	53.6 ± 8.84	53.1 ± 8.99	0.6
TAC (µmol/L)	5.69 ± 1.14	5.46 ± 1.24	5.2 ± 1.25	0.05*

[Table/Fig- 4]: Levels of serum oxidative markers and activities of antioxidant enzymes in IGE patients with different time duration of AED treatment. Note: p-value <0.05* is significant
0-≤1 year - Patients who were on AED treatment for less than 1 year
1-≤5 years- Patients who were on AED treatment for 1-5 years duration
>5 years- Patients who were on AED treatment for more than 5 years

Variable	Treated Patients (n = 235)	Untreated Patients (n = 75)	p-value
NO (µmol/mL)	8.55 ± 2.0	7.21 ± 1.86	<0.0001*
MDA (nmol/mL)	4.31 ± 1.65	3.82 ± 1.38	0.02*
SOD (U/gHb)	606 ± 88	676.9 ± 82.1	<0.0001*
CAT (k/ml)	5.7 ± 1.24	6.21 ± 1.1	0.001*
GPx (U/gHb)	53.8 ± 8.85	55.9 ± 6.81	<0.06
TAC (µmol/L)	5.47 ± 1.23	5.82 ± 0.96	<0.02*

[Table/Fig- 5]: Comparison between oxidative stress markers, antioxidant enzyme activities in untreated and treated (with AEDs) patients of GGE. Note: p-value <0.05* is significant

53.8±8.85 U/gHb and 843±95.2, 9.39±1.33, 76.7±11.7 respectively) ($p < 0.001$). The levels of oxidative markers and antioxidant enzyme activities in IGE patients with AED treatment duration (≤ 1 year, 1- ≤ 5 years and > 5 years) have been shown in [Table/Fig-4].

Statistically significant difference in the levels of NO, MDA, activities of SOD, CAT and TAC were observed among patient subgroups who were on AEDs treatment for different time periods ($p = 0.02$, $p = 0.01$, $p = 0.001$, $p = 0.01$, $p = 0.05$ respectively) whereas no significant difference was noted in the activity of GPx between these subgroups. Finally, an attempt was made to compare the oxidative markers, antioxidant enzymes in IGE treated patients (AEDs) with untreated patients and the results have been depicted in [Table/Fig-5].

Significantly increased levels of NO, MDA, decreased activities of antioxidant enzymes (SOD, CAT) and TAC were observed in IGE treated patients compared to untreated patients ($p < 0.001$, $p = 0.02$, $p < 0.001$, $p = 0.001$, and $p = 0.02$ respectively).

DISCUSSION

Many studies have suggested that the serum electrolytes, trace elements and lipid peroxidation, due to oxidative stress are causally involved in some epilepsy types and their seizure recurrence [21]. The brain contains high concentration of PUFA that are more prone to lipid peroxidation. Hence, the assessment of MDA levels in biological materials can be used as an important indicator of lipid peroxidation in vitro and in vivo for various diseases. In the present study, we found significantly higher levels of MDA in IGE patients compared to controls. Our results are in accordance with many previous studies where a higher level of MDA has been reported to be associated with epilepsy [22-24]. Conversely, no difference [25] or significantly decreased [26] levels of MDA have been reported in epilepsy patients. It was also found that significantly increased MDA levels in treated (with AEDs) compared to untreated IGE patients suggest that the additional oxidative stress was induced by AEDs, thereby causing seizure recurrence as well as drug intolerance.

It has been suggested that in CNS, NO plays a significant role in epilepsy and epileptogenesis because it acts as a secondary messenger, neuromodulator and neurotransmitter [27]. In the

present study, NO levels were found to be significantly high in patients when compared to controls. Our results are in agreement with many other studies, where significantly elevated NO levels have been reported to be associated with the disease [25,28]. Similarly, significantly elevated serum NO levels were observed in treated patients compared to untreated patients denoting that additional oxidative stress was induced by AEDs which results in seizure recurrence and drug intractability.

The superoxide dismutases play a crucial role in eliminating superoxide anion radicals ($O_2^{\cdot-}$) generated from extracellular stimulants, including ionizing radiation and oxidative insults together with produced in mitochondrial matrix [29]. In the present study, we found a significant decrease in SOD activity in patients as compared to controls. Our results are in accordance with earlier studies where a significantly lower activity has been found in patients with GGE [30]. Conversely, increased levels of SOD have been noted in drug naive patients with epilepsy [22] and unchanged SOD levels in other studies [28]. In order to maintain the proper functioning of Cu-Zn SOD1 enzyme, the Cu-Zn ratio is important rather than absolute amount of Cu or Zn alone [31]. In our previous study, we found significantly low levels of serum zinc and increased Cu/Zn ratio in GGE as well as in IGE patients [32]. In addition, significantly lower activities of SOD were observed in treated patients of epilepsy compared to untreated patients suggesting that additional oxidative stress induced by AEDs may interfere with the pathogenesis of epilepsy. Hence, in addition to high Cu/Zn ratio, low levels of Zn and SOD reflects SOD function which may lead to ineffectual scavenging of ROS and resulting in seizure recurrence.

Our study showed significant lower activities of CAT in GGE patients compared to controls. Our results are in agreement with the earlier studies, where significant low levels were claimed in patients compared to controls [22,33]. Also, the CAT levels were found to be significantly low in treated patients compared to untreated patients. The low levels of CAT can be attributed to inactivation of enzyme by superoxide anions and thereby inducing additional oxidative stress indirectly which results in seizure promotion and drug intractability as well.

In the present study, the erythrocyte GPx activity was found to be significantly low in patients when compared to controls. Our results are in accordance with Saad K et al., who reported both Selenium (Se) and GPx activity to be reduced in children with intractable seizures [34]. Significantly lower activity of GPx was observed in treated patients as compared to untreated patients suggest that increased oxidative stress induced by AEDs may interfere in the pathogenesis of seizures and causing unresponsiveness to AEDs.

TAC can be considered as a cumulative index of complete antioxidant capacity. In the present study, the level of TAC were significantly decreased in IGE patients compared to controls and in treated patients compared to untreated patients suggesting that lower antioxidant enzymes activity may also contribute to the ongoing process of epileptogenesis.

Accumulating evidence suggests that the chronic usage of first-choice AEDs such as valproic acid, phenytoin and carbamazepine increases free radical formation followed by oxidative damage in neurons [7]. But AEDs such as zonisamide and topiramate have antioxidant effects by scavenging hydroxyl radicals and NO in experimental conditions [4]. The possible mechanism behind oxidative stress induced by AEDs is that these drugs are metabolized to reactive epoxide intermediates, which may covalently bind to biomolecules such as lipids, proteins etc., and induce structural and functional impairments of the cell rather than having a neuroprotective effect [35]. In the present study, a significant difference in the levels of NO, MDA, decreased activity of SOD and CAT were noticed between the patient subgroups of AED treatment duration viz., ≤ 1 year, 1- ≤ 5 years and > 5 years. A significant decrease in the activities of SOD, CAT and TAC and increased level of NO and MDA in treated patients

compared to untreated patients of GGE signify the role of AEDs in the induction of oxidative stress, utilization of antioxidant enzymes and thereby reducing the antioxidant capacity that may result in seizure recurrence and drug unresponsiveness. Our results are in accordance with previous study in which the decreased activities of antioxidant enzymes and also a positive effect of AEDs on oxidative stress were noticed [36]. Our results suggest that chronic usage of AEDs may trigger oxygen-dependent tissue injury that lead to additional oxidative stress, which may be implicated in the seizure generation, progression and drug intractability.

LIMITATION

Further studies are required to establish the role of individual AEDs in the development of oxidative stress. The clinical relevance of the study has to be evaluated.

CONCLUSION

The present study demonstrates that the epilepsy seems to cause the elevation of NO and MDA levels that reflects the oxidative stress in GGE patients. The pro-oxidative effects of AEDs might lead to additional oxidative stress, increased seizure activity resulting in loss of their efficiency and drug intractability. The concomitant use of antioxidants along with conventional AEDs could have a beneficial effect.

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REFERENCES

- Hesdorffer DC, Logroscino G, Benn EK, Katri N, Cascino G, Hauser WA. Estimating risk for developing epilepsy: a population based study in Rochester, Minnesota. *Neurology*. 2011;76:23–27.
- Panayiotopoulos CP. Idiopathic generalized epilepsies: A review and modern approach. *Epilepsia*. 2005;46: 1–6.
- Deepa D, Jayakumari N, Thomas SV. Oxidative stress is increased in women with epilepsy: Is it a potential mechanism of anti-epileptic drug-induced teratogenesis? *Ann Indian Acad Neurol*. 2012;15:281–86.
- Shin EJ, Jeong JH, Chung YH, Kim WK, Ko KH, Bach JH, et al. Role of oxidative stress in epileptic seizures. *Neurochem Int*. 2011;59:122–37.
- Ashrafi MR, Shams S, Nouri M, Mohseni M, Shabani R, Yekaninejad MS, et al. A probable causative factor for an old problem: selenium and glutathione peroxidase appear to play important roles in epilepsy pathogenesis. *Epilepsia*. 2007;48:1750–55.
- Nita M and Grzybowski A. The role of the reactive oxygen species and oxidative stress in the pathomechanism of the age-related ocular diseases and other pathologies of the anterior and posterior eye segments in adults. *Oxid Med Cell Longev*. 2016;2016:3164734–57.
- Martinc B, Grabnar I, Vovk T. The role of reactive species in epileptogenesis and influence of antiepileptic drug therapy on oxidative stress. *Curr Neuropharmacol*. 2012;10(4):328–43.
- Shichin M. The role of lipid peroxidation in neurological disorders. *J Clin Biochem Nutr*. 2014;54(3):151–60.
- Santoshi RG, Adinath NS, Prabhakar MK. Study of lipid peroxidation and total antioxidant capacity in epileptic patients. *Int J of Curr Res*. 2012;4(9):12–14.
- Cardenas-Rodriguez N, Huerta-Gertrudis L, Rivera- Espinosa, Montesinos-Correa H, Bandala C, Carmona-Aparicio L, et al. Role of oxidative stress in refractory epilepsy: evidence in patients and experimental models. *Int J of Mol Sc*. 2013;14:1455–76.
- Ho YS, Xiong Y, Ma W, Spector A, Ho DS. Mice lacking catalase develop normally but show differential sensitivity to oxidant tissue injury. *J Biol Chem*. 2004;279(31):32804–12
- Commission on classification and terminology of the International League Against Epilepsy. Proposal for revised classification of epilepsies and epileptic syndromes. *Epilepsia*. 1989;30:389–99.
- Cutting S, Lauchheimer A, Barr W, Devinsky O. Adult-onset idiopathic epilepsy: clinical and behavioural features. *Epilepsia*. 2001;42:1395–98.
- Lepoivre M, Chenais B, Yapo A, Lemaire G, Thelander, Tenu JP. Alterations of ribonucleotide reductase activity following induction of the nitrite-generating pathway in adenocarcinoma cells. *J Biol Chem*. 1990;265:14143–49.
- Buege JA, Aust SD. Microsomal lipid peroxidation. *Methods Enzymol*. 1978;105:302–10
- Benzie IF, Strain JJ. The ferric reducing ability of plasma (FRAP) as a measure of “antioxidant power”. The FRAP assay. *Anal Biochem*. 1996;239:70–76.
- Rotruck JT, Pope AL, Ganther HE, Swanson AB, Hafeman DG, Hoekstra WG. Selenium: biochemical role as a component of glutathione peroxidase. *Science*. 1973;179:588–90.
- Marklund SL, Marklund G. Involvement of superoxide Anion Radical in the auto oxidation of pyrogallol and a convenient assay for superoxide dismutase. *European J Biochem*. 1974;47:469–74.
- Aebi H. Catalase in vitro. *Methods Enzymol*. 1984;105:121–26.
- Marini C, King MA, Archer JS, Newton MR, Berkovic SR. Idiopathic generalised epilepsy of adult onset: clinical syndromes and genetics. *J Neurol Neurosurg Psychiatry*. 2003;74:192–96.
- Wojciak RW, Mojs E, Stanislawski-Kubiak M, Samborski W. The serum zinc, copper, iron, and chromium concentrations in epileptic children. *Epilepsy Res*. 2013;104:40–44.
- Guler SK, Aytac B, Durak ZE, Cokal BG, Gunes N, Durak I, et al. Antioxidative-oxidative balance in epilepsy patients on antiepileptic therapy: a prospective case-control study. *Neurolog Sci*. 2016;37(5):763–67.
- Çevik MU, Varol S, Yücel Y, Akil E, Çelepkolu T, Arkanolu A, et al. Serum paraoxonase-1 activities and malondialdehyde levels in patients with epilepsy. *Dicle Med J*. 2012;39(4):557–60.
- Pandey MK, Mitra P, Maheshwari PK. The Lipid peroxidation product as a marker of oxidative stress in epilepsy. *J Clin Diagn Res*. 2012;6(4):90–92.
- Ebru A, Ayse S, Bahadır O, Serdar OH, Aysima O, Nese K, et al. Effects of epilepsy and antiepileptic drugs on nitric oxide, lipid peroxidation and xanthine oxidase system in children with idiopathic epilepsy. *Seizure*. 2011;20(2):138–42.
- Yis U, Seckin E, Kurul S, Kuralay F, Dirik E. Effects of epilepsy and VPA on oxidant status in children with idiopathic epilepsy. *Epilepsy Res*. 2009;84:232–37.
- Banach M, Piskorska B, Czuczwar SJ, Borowicz KK. Nitric oxide, epileptic seizures, and action of antiepileptic drugs. *CNS Neurol Disord Drug Targets*. 2011;10:808–19.
- Peker E, Oktar S, Ari M, Kozan R, Dogan M, Cagan E, et al. Nitric oxide, lipid peroxidation, and antioxidant enzyme levels in epileptic children using valproic acid. *Brain Res*. 2009;1297:194–97.
- Miao L, St. Clair DK. Regulation of superoxide dismutase genes: implications in disease. *Free Radic Biol Med*. 2009;47(4):344–56.
- Nemade ST, Melinkeri RR. Oxidative and antioxidant status in epilepsy. *Pravara Med Rev*. 2010;2:8–10.
- Harris ED. Copper homeostasis: the role of cellular transporters. *Nutr Rev*. 2001;59:281–85.
- Prasad DKV, Shaheen U, Satyanarayana U, Surya Prabha T, Jyothy A, Munshi A. Association of serum trace elements and minerals with genetic generalized epilepsy and idiopathic intractable epilepsy. *Neurochem Res*. 2014;39:2370–76.
- Ogunro PS, Mustapha AF, Salau AA. Lipid peroxidation and antioxidant status in patients with primary generalized epilepsy. *Arch Appl Sci*. 2013;5:68–74.
- Saad K, Hammad E, Hassan AF, Badry R. Trace element, oxidant, and antioxidant enzyme values in blood of children with refractory epilepsy. *Int J of Neurosci*. 2014;124(3):181–86.
- Grewal GK, Kukal S, Kanojia N, Saso L, Kukreti S, Kukreti R. Effect of oxidative stress on ABC transporters: Contribution to epilepsy pharmacoresistance. *Molecules*. 2017;22(3):365–78.
- Ercegovac M, Jovic N, Simic T, Bumbasirevic LB, Sokic D, Radojevic AR. Antiepileptic drugs affect protein, lipid and DNA oxidative damage and antioxidant defense in patients with epilepsy. *J Med Biochem*. 2013;32:121–30.

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